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Research Article

In Vitro Investigation of a Terbinafine Impregnated Subcutaneous Implant for Veterinary Use

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A terbinafine impregnated subcutaneous implant was evaluated to determine if drug was released into isotonic saline over the course of 6 months at two different temperatures, 37°C and 4°C. These temperatures were chosen to simulate the nonhibernating (37°C) and hibernating body (4°C) temperatures of little brown bats (Myotis lucifugus). Insectivorous bats of North America, including little brown bats, have been devastated by white nose syndrome, a fungal infection caused by Geomyces destructans. No treatments exist for bats infected with G. destructans. Implants were placed into isotonic saline; samples were collected once per week and analyzed with HPLC to determine terbinafine concentrations. The mean amount of terbinafine released weekly across the 28 weeks was approximately 1.7 µg at 4°C and 4.3 µg at 37°C. Although significant differences in the amount released did occur at some time points, these differences were not consistently greater or less at either of the temperatures. This study showed that terbinafine was released from an impregnated implant over the course of 6 months at concentrations ranging from 0.02 to 0.06 µg/mL depending on temperature, which may be appropriate for little brown bats (Myotis lucifugus) infected with Geomyces destructans, the etiologic agent of white nose syndrome.

1. Introduction

Treatment of systemic fungal infections often requires from weeks to months of drug therapy. Consistently medicating companion animals for this length of time can be difficult and even more so with animals that become stressed with handling, such as wildlife or exotic pets. Nondomesticated animals are susceptible to stress from repeated handling and restraint, and stress can lead to the death of hospitalized wildlife or exotic pets [1]. Stress, including that associated with handling in animals, has also been shown to lead to immunosuppression and increased susceptibility to disease. Therefore, stress associated with repeated handling for treatment of an infection could inhibit an animal’s ability to mount an appropriate immune response [2–4].

White nose syndrome, caused by the fungus Geomyces destructans, is an infection that affects insectivororous bats of North America [5–7]. Extremely high mortality rates are associated with infection and to date, no effective treatments have been found. If numbers of bats continue to decline and approach extinction, members of various affected species may need to be captured and kept in captivity in order to preserve genetic diversity for eventual release and repopulation. Animals may be infected with G. destructans when captured and need antifungal treatment that is long acting and which would require limited handling of the animal.

Terbinafine is a fungicidal medication that inhibits the synthesis of ergosterol which is an essential component of fungal walls. Although no studies examining this drug in bats have been published, studies have been performed in other animal species [8–12]. Geomyces pannorum, a fungus that is closely related to G. destructans, can cause infection in humans and is susceptible to terbinafine [13, 14]; no published reports regarding the sensitivity of G. destructans to terbinafine are available. Terbinafine has also been useful
in other refractory mycotic infections in humans [15]. *Geomyces destructans* has been shown to be susceptible to other antifungal agents in vitro including fluconazole, but terbinafine has a better safety profile than many other commonly used antifungal medications [16–18].

The goal of this study was to investigate a terbinafine impregnated implant designed for subcutaneous placement over the dorsum of bats infected with *G. destructans*; the in vitro release of terbinafine from the implant was evaluated at two different temperatures, 4°C and 37°C, over the course of approximately 6 months. The two temperatures were chosen because they are similar to the body temperatures of hibernating (4°C) and nonhibernating (37°C) bats. This initial trial was designed to determine if terbinafine would release from the implant over the course of many months without degradation of the implant in an in vitro setting.

### 2. Materials and Methods

Implants were constructed by Melatek, LLC (Madison, WI, USA) based on protocols used to make Ferretonin implants. These implants are stable for approximately 5 years if kept at 4°C (T. Cairns, pers. comm.). Briefly, terbinafine HCl (Sigma-Aldrich, Co., St. Louis, MO, USA) was mixed with medical grade elastomer to a calculated concentration so that each implant would contain 0.5 mg of terbinafine. The mixture was placed into a mold where it cured and was then cut into individual implants. Each implant was approximately the size of a passive integrated transponder (PIT) tag (microchip) as shown in Figure 1. Cured medical grade elastomer is dimensionally and thermally stable, resistant to oxidation and sunlight, and does not become grade elastomer is dimensionally and thermally stable, resistant to oxidation and sunlight, and does not become

![Image 1: The terbinafine impregnated implant (arrow) is shown next to a PIT tag (microchip) and penny.](image)

For in vitro analysis, implants were individually placed into 25 mL of isotonic saline in glass containers. Five implants were kept at 4°C and five were kept at 37°C. Every 7 days, the solution in each container was mixed to ensure homogeneity and 350 µL was then removed and placed into a cryo-tube for analysis. Following sample removal, isotonic saline (350 µL) was added to the container so the volume was kept consistent. Samples were collected for a total of 28 weeks and were kept in a −80°C freezer until analysis.

Saline samples were analyzed using HPLC with ultraviolet absorption. The system consisted of a 2695 separations module, a 2487 absorbance detector (Waters, Milford, MA, USA). Terbinafine was extracted from saline samples using a hexane extraction and was separated on a Symmetry Shield C18 (4.6 x 100 mm, 5 µm) column with a guard column. The mobile phase was a mixture of (A) 20 mM phosphoric acid with 0.1% triethylamine adjusted to pH 3.0 and (B) acetonitrile (65:35). The flow rate was 1.1 mL/min and the column temperature ambient. Absorbance was measured at 224 nm.

Standard curves for analysis were prepared by fortifying saline with terbinafine to produce a linear concentration range of 5–1500 ng/mL. Average recovery for terbinafine was

<table>
<thead>
<tr>
<th>Weeks after initial placement into saline</th>
<th>4°C Mean (SD)</th>
<th>37°C Mean (SD)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>3.4 (0.6)</td>
<td>4.8 (0.2)</td>
<td>&lt;0.01*</td>
</tr>
<tr>
<td>2</td>
<td>20.3 (12.7)</td>
<td>5.2 (4.6)</td>
<td>0.04*</td>
</tr>
<tr>
<td>3</td>
<td>19.9 (21.2)</td>
<td>−8.0 (6.1)</td>
<td>0.02*</td>
</tr>
<tr>
<td>4</td>
<td>3.7 (15.1)</td>
<td>20.3 (21.3)</td>
<td>0.19</td>
</tr>
<tr>
<td>5</td>
<td>5.7 (1.9)</td>
<td>−3.2 (23.0)</td>
<td>0.75</td>
</tr>
<tr>
<td>6</td>
<td>−2.3 (10.6)</td>
<td>12.6 (10.8)</td>
<td>0.06</td>
</tr>
<tr>
<td>7</td>
<td>−20.2 (55.6)</td>
<td>−1.6 (21.0)</td>
<td>0.70</td>
</tr>
<tr>
<td>8</td>
<td>6.0 (54.3)</td>
<td>14.2 (7.5)</td>
<td>0.17</td>
</tr>
<tr>
<td>9</td>
<td>10.9 (14.7)</td>
<td>14.4 (12.4)</td>
<td>0.70</td>
</tr>
<tr>
<td>10</td>
<td>−0.4 (8.9)</td>
<td>4.2 (33.2)</td>
<td>0.60</td>
</tr>
<tr>
<td>11</td>
<td>−2.1 (6.0)</td>
<td>5.5 (6.3)</td>
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</tr>
<tr>
<td>12</td>
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<td>0.60</td>
</tr>
<tr>
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<td>6.3 (4.7)</td>
<td>0.88</td>
</tr>
<tr>
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<td>−4.7 (9.3)</td>
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<tr>
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<td>−3.7 (8.5)</td>
<td>0.86</td>
</tr>
<tr>
<td>16</td>
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<td>0.44</td>
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<tr>
<td>17</td>
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<td>11.0 (5.5)</td>
<td>&lt;0.01*</td>
</tr>
<tr>
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<td>2.9 (5.6)</td>
<td>0.19</td>
</tr>
<tr>
<td>19</td>
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<td>13.4 (6.9)</td>
<td>0.78</td>
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<tr>
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<tr>
<td>21</td>
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<td>0.10</td>
</tr>
<tr>
<td>22</td>
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<td>15.0 (27.7)</td>
<td>0.32</td>
</tr>
<tr>
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<td>0.35</td>
</tr>
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<td>−9.5 (5.0)</td>
<td>0.45</td>
</tr>
<tr>
<td>25</td>
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<td>10.2 (4.1)</td>
<td>0.03*</td>
</tr>
<tr>
<td>26</td>
<td>−6.3 (2.5)</td>
<td>−2.6 (3.3)</td>
<td>0.08</td>
</tr>
<tr>
<td>27</td>
<td>−1.9 (4.9)</td>
<td>6.7 (6.4)</td>
<td>0.04*</td>
</tr>
</tbody>
</table>

*P value less than 0.05 indicates a significant difference.*
95% while intra- and interassay variability were less than 10%. The lower limit of quantification was 5 ng/mL.

Following HPLC analysis, the amount of terbinafine released by each implant during each interval was calculated. The mean release of terbinafine with standard deviations was calculated for the different temperatures at each time point. Data was tested for normalcy with a Bartlett’s test for inequality of variances. If the values were normally distributed, a t-test was performed to determine if a significant difference in amount of terbinafine released was present at the two temperatures. If the data was not normally distributed, a Mann-Whitney/Wilcoxon two-sample test was used to determine if differences existed. Significance was set at \( P < 0.05 \) and analysis was performed with EpilInfo (CDC, Atlanta, GA, USA).

3. Results

Samples were collected and analyzed with HPLC for a total of 28 weeks after initial placement into isotonic saline. A sample was not collected during week 23. The mean amount released from the implants at 37°C was significantly greater than 4°C at the 1 \( P < 0.01 \), 17 \( P < 0.01 \), 26 \( P = 0.03 \), and 28 \( P = 0.04 \) week time points; the amount released from implants at 4°C was greater than 37°C at the 2 \( P = 0.04 \) and 3 \( P = 0.02 \) week time points. The mean amount of terbinafine released weekly across the 28 weeks was approximately 1.7 \( \mu g \) at 4°C and 4.3 \( \mu g \) at 37°C.

If terbinafine was released from implants at different rates at the different temperatures, excessively high concentrations may be reached in nonhibernating bats or suboptimal concentrations may be reached in hibernating bats. This study, however, further studies are needed to determine the MIC of \( G.\ destructans \). Additionally, initial clinical trials in little brown bats are currently being performed (M. Souza, pers. comm.). Implants were placed subcutaneously over

4. Discussion

The implant was evaluated at two different temperatures because of the differing rates of metabolism between hibernating and nonhibernating bats [19]. It was hypothesized that more terbinafine would be released at 37°C than at 4°C. However, further studies are needed to determine the MIC of \( G.\ destructans \). Additionally, initial clinical trials in little brown bats are currently being performed (M. Souza, pers. comm.). Implants were placed subcutaneously over

Figure 2: Terbinafine impregnated implants were placed into isotonic saline at 4°C (\( n = 5 \)) and 37°C (\( n = 5 \)). Samples were collected every 7 days and terbinafine concentrations were determined with HPLC. A sample was not collected during week 23. The mean amount (\( \mu g \)) of terbinafine released (\( \pm \)SD) at the different temperatures is shown over the course of 6 months.
the dorsum of bats infected with *G. destructans* and safety and efficacy of the implants will be determined. Results are not yet available, but skin samples will be evaluated with HPLC to determine terbinafine concentrations.

5. Conclusions

In conclusion, terbinafine was released from the implant over the course of 6 months with no consistent significant differences at two different temperatures, 37°C and 4°C. However, without in vivo absorption, metabolism and clearance, it is difficult to know whether this implant will release therapeutic amounts of terbinafine in *G. destructans* infected bats. This research was the first step to determine if terbinafine would release from the implant over an extended period of time and what amounts might be released. Future research will need to examine the implants in animals to determine the concentration of systemic terbinafine over time. Following further investigation, this implant may provide a long term treatment for *G. destructans* infected bats that requires handling only once at the beginning of treatment.

References


