Hemiparasitic Castilleja promotes plant diversity in an alpine meadow ecosystem

Johannah Reed
jreed41@utk.edu

Follow this and additional works at: https://trace.tennessee.edu/utk_chanhonoproj

Part of the Biology Commons, Other Ecology and Evolutionary Biology Commons, Plant Biology Commons, and the Terrestrial and Aquatic Ecology Commons

Recommended Citation

This Dissertation/Thesis is brought to you for free and open access by the University of Tennessee Honors Program at Trace: Tennessee Research and Creative Exchange. It has been accepted for inclusion in University of Tennessee Honors Thesis Projects by an authorized administrator of Trace: Tennessee Research and Creative Exchange. For more information, please contact trace@utk.edu.
Hemiparasitic Castilleja promotes plant diversity in an alpine meadow ecosystem

By Johannah Reed

Mentor: Dr. Nate Sanders
ABSTRACT

There is a long history in ecology of examining how biotic interactions such as competition, predation, and mutualism influence the structure and dynamics of natural communities. However, few studies to date have experimentally assessed the role of hemiparasitic plants, which are commonly found in a variety of ecosystems, as a structuring force of plant communities. Hemiparasitic plants have the potential to shape plant communities because of their ability to photosynthesize and parasitize host plants and because of their abundance in a variety of natural and managed ecosystems. I conducted a study that focused on the impacts of hemiparasitic species in the genus *Castilleja*, which are ubiquitous in montane meadows in the Rocky Mountain West. This study employed a manipulative field experiment in which *Castilleja* was removed from intact plant communities to examine the relationship between the presence of *Castilleja* and a suite of community-level properties. The experiment showed that evenness was 32% lower, diversity was 29% lower, and Shannon entropy was 32% lower in plots from which *Castilleja* was removed relative to plots in which *Castilleja* was present. This suggests that the presence of *Castilleja* promotes diversity in a montane plant community. This research demonstrates that hemiparasites have the potential to shape plant communities in alpine ecosystems, but more research should be done to determine the extent of their effects on community structure and ecosystem function in natural and manipulated systems.
INTRODUCTION

Plant community ecologists have examined the effects of competition and herbivory on plant community structure for at least decades, but they have generally ignored the impact parasitic or hemiparasitic plants might have on community composition (Spasojevic and Suding 2010; Bardgett et al. 2006). Parasitic plants are comprised of two main groups, holoparasites (plants that are completely parasitic and contain virtually no chlorophyll) and hemiparasites (plants that are both parasitic and photosynthetic), and each group has the potential to play a major role in shaping plant community structure (Press and Phoenix 2005). Globally there are over 3000 parasitic flowering plant species, and most plant assemblages contain one or more parasitic plant species (Marvier 1996; Bardgett et al. 2006; Quested 2008). Parasitic plants are also represented in every major ecosystem (Adler 2002). Given their ubiquity and diversity, it is surprising that so few studies have examined whether they impact plant community composition.

Hemiparasitic plants may exert strong indirect effects on plant community structure by directly altering the performance of their hosts. Though few studies have examined community-level impacts of hemiparasitic plants (see Bardgett et al. 2006; Phoenix and Press 2005), several experiments on parasite-host interactions have found that hemiparasites can have detrimental effects on their hosts and that hosts have varying positive effects on their parasite (Adler 2003; Pennings and Simpson 2008). Hemiparasitic plants obtain some resources, including water, fixed carbon compounds, nitrogen, etc., from their host plants, but they also compete with their hosts for light, water and soil nutrients (Pennings and Callaway 2002; Press and Phoenix 2005; Marvier 1996; Borowicz and Armstrong 2011). Hemiparasitic plants have the potential to determine the structure and productivity of the plant communities in which they are present, as their parasitism can decrease the growth of host species, which might indirectly affect other
plants by altering the competitive balance between hosts and non-hosts species in a community (Press and Phoenix 2005; Quested, et al. 2002). Put another way, when hemiparasites have detrimental effects on their hosts, this can allow non-host plants to gain a competitive advantage and become more abundant in the community (Bardgett et al. 2006; Marvier 1998). Finally, the effects of hemiparasitic plants on community biomass or diversity depends on whether their host is a dominant or subordinate species in the community. If a host is a dominant species, then hemiparasitic species might promote diversity and decrease ANPP when compared to when it parasitizes a subordinate host (Bardgett et al. 2006).

Soil nutrient availability, the density of parasitic plants within the community, and the parasitic plants’ host preference can all drive changes in plant community composition (Spasojevic and Suding 2010; Quested 2008). Most parasitic plants are thought to be generalists, meaning that they parasitize and exploit resources from many different plant species, and Castilleja spp. are even thought to parasitize over 100 different hosts (Press 1998). This could drive changes in competitive interactions among hosts or create other indirect effects (Marvier 1998). It is common, however, for parasitic plants to behave more as specialists because they show various levels of host preference (Press and Phoenix 2005).

Castilleja species are facultative hemiparasites of plant roots, from which they obtain a portion of their required nutrients, including water, nitrogen, carbon compounds, and minerals from their host (Bardgett et al. 2006; Press and Phoenix 2005; Cameron et al. 2009; Quested 2008; Marvier 1996). Hemiparasitic plants such as Castilleja species can “forage” by selectively growing in a specific direction (towards or away from host plants) or by selectively penetrating a host’s roots upon contact (Pennings and Callaway 2002). It is difficult to determine the host for most Castilleja individuals, but Castilleja species can parasitize a wide range of plant species in
montane and alpine communities (Spasojevic and Suding 2010), including deeply rooted
sagebrush, fast growing grasses, lupines, and other plants (Darrow 2006). Adler (2003) found
that *Castilleja indivisa* benefits more from parasitizing lupines, which are nitrogen fixers,
suggesting that some hosts have stronger positive effects on the hemiparasite’s productivity than
others (Adler 2003; Pennings and Simpson 2008).

Although *Castilleja* species are common in alpine systems in Colorado and the Rocky
Mountains (Appendix 1), their community-level effects have not been widely studied. One
exception is the work of Spasojevic and Suding (2010), who conducted an experiment in the
Front Range of the Colorado Rocky Mountains focusing on the effects of *Castilleja occidentalis*
on nutrient dynamics and diversity-productivity relationships. They found that long-lived
hemiparasitic plants may increase the availability of nitrogen in nutrient poor systems through
their nutrient rich litter. This positive nutrient effect could significantly increase productivity and
slightly increase diversity. To my knowledge, this is the only study that shows the effects of
hemiparasitism on community structure and driving ecosystem processes in Rocky Mountain
ecosystems.

In my study, I employed a manipulative experiment to examine the relationship between
the presence of *Castilleja* spp. and a suite of community-level variables. In particular, I asked:
(1) Do *Castilleja* spp. affect the structure of plant communities by altering species richness,
evenness, and diversity? (2) How do *Castilleja* spp. affect the structure of plant communities? I
predicted that if *Castilleja* species have an effect on plant communities, *Castilleja* presence
would promote species richness, evenness, and diversity. Overall, with this research I aim to
better understand the roles of parasitic plants in shaping community structure in alpine systems.
METHODS

My work was conducted near the Rocky Mountain Biological Laboratory (RMBL) (latitude 38° 96’ N, longitude 106 ° 98’ W). The experimental site was located at approximately 3000 meters in elevation in a dry montane meadow approximately 1-km east of Gothic, CO. The dominant vegetation at the site consisted of herbaceous perennials, aspen trees, and a mixture of grasses. There were about fifty plant species represented across the study site; common plant species at the site included *Populus tremuloides, Achillea millefolium, Potentilla hippiana, Mahonia repens, Lomatium dissectum, Ipomopsis aggregata, Phleum praetense* (Appendix 1).

In July 2011, I established 18 1-m² plots with a 0.5 × 0.5 m buffer zone to determine if *Castilleja* influences the structure and diversity of plant communities. Each plot was at least 1-2 m from the nearest adjacent plots, but the exact distance was not measured, as it depended on the topography and vegetation of the area, as well as the abundance of *Castilleja*. I implemented three treatments: a control plot in which nothing was manipulated; a *Castilleja*-removal plot, in which *Castilleja* was removed by clipping to ground level in each plot (including the 0.5 × 0.5 m buffer zone) each week during the growing season (primarily July 2011); and a random-biomass removal plot, in which a comparable amount of biomass (compared to the *Castilleja*-removal plots) was removed. I included a random-biomass removal plot in each block to account for the removal of *Castilleja* biomass and the impacts that removing biomass in general could have on plant community structure (Symstad and Tilman 2001). Biomass removed from the *Castilleja*-removal plots and random-biomass removal plots was collected in paper bags and then dried for approximately 72 hours at 70°C in a static air oven. Collected biomass was weighed to the nearest 0.01 gram.
To determine if all plots were similar in their abiotic pressures and properties, soil moisture was measured for all plots periodically using a soil moisture probe with all measurements taken on clear days between 11 am and 1 pm. Two measurements were taken in each plot on each measurement occasion throughout the peak growing time for *Castilleja*.

To determine the effects of *Castilleja* on plant community structure, I measured the richness, evenness, and diversity of the existing plant community on three occasions (June 28th, July 14th and July 28th) during the 2011 growing season in each of the 18 1-m² plots; however, I used only the data from the last observation at the end of the growing season to measure the effects of *Castilleja* on the community-level variables. Richness was measured in all plots on each date by identifying and recording each plant species present in the plots (Darrow 2006). Evenness was measured by estimating the relative abundance of each species based on percent foliar cover at five percent intervals and using the Inverse Simpson’s Index

\[
\text{Evenness} = \left(\frac{1}{\sum p_i \ln p_i}\right) \frac{1}{S}
\]

where \( p_i \) is the proportion of each species and \( S \) is the total number of species. Shannon’s Diversity \( H' = -\sum p_i \ln p_i \) and Shannon’s Entropy \( \text{Entropy} = e^{-H'} \) were also calculated. Note that in all cases the response variable did not include *Castilleja* presence or abundance; instead, the focus of these calculations was on the diversity of the rest of the plant community.

For each response variable, I used a separate one-way Analysis of Variance model with treatment being “removal” at three levels: (1) *Castilleja* removal, (2) random biomass removal, and (3) control. I also used Tukey-Kramer HSD to determine whether treatments differed from each other for each of the response variables at \( \alpha = 0.05 \). All data met the assumptions of normality prior to analysis.
RESULTS

Prior to plant removals in 2011, no diversity measures differed among treatments (P > 0.667 in all cases), and plots did not differ in soil moisture (P = 0.971; F = 0.030). After plant removals, richness still did not differ among treatments (Table 1; Figure 1A), but evenness (Inverse Simpson’s Index) and diversity (Shannon’s Diversity and Shannon’s Entropy) did significantly differ (Table 1). In particular, evenness was 32% lower in *Castilleja* removal plots than in control plots (Figure 1B; F = 18.56; P < 0.0001; Table 1), and diversity was 29% lower in *Castilleja* removal plots than in control plots (Figure 1C; F = 8.14; P < 0.004; Table 1). Shannon’s Entropy was marginally significant (P = 0.070, F = 3.18), showing that *Castilleja* removal plots had 32% lower Shannon’s entropy than control plots (Figure 1D). Finally, Shannon’s diversity and entropy did not differ between *Castilleja* removal and random biomass removal plots; random biomass removal plots did not differ from control plots either in terms of evenness, Shannon’s diversity, and Shannon’s entropy (Table 1; Figure 1C, D).
TABLE 1. ANOVA results testing for the effects of *Castilleja* removal on Simpson’s evenness, Shannon’s diversity, and Shannon’s entropy after a growing season of experimental treatments. Significant (P < 0.05) and marginally significant variables (P < 0.07) are in bold.
**FIGURE 1.** Mean (± SE) values for plant community diversity measures as they are affected by the presence of *Castilleja* spp. Richness or number of species in plots (A), Inverse Simpson’s Evenness Index (B), Shannon’s Diversity Index (C), and Shannon’s Entropy (D) for *Castilleja* removal plots, control plots, and random biomass removal plots. Values with different letters are significantly different at the P <0.05 level, except for D, in which P<0.07 is considered marginally significant.
DISCUSSION

The most salient result of this study was that the presence of the hemiparasite *Castilleja* altered alpine plant community structure over the course of a single growing season. More specifically, when *Castilleja* was present, plant community evenness, diversity, and entropy were all higher than when *Castilleja* species were experimentally removed (Figure 1B, 1C, 1D). Species richness was not affected by the presence of *Castilleja* in the experiment (Figure 1A). Taken together, the results of this study suggest that the presence of the hemiparasite *Castilleja* promotes shifts in community structure and overall higher diversity in this alpine system.

Hemiparasitism by *Castilleja* promoted evenness and diversity while leaving species richness unchanged. It is not surprising that richness did not differ among treatments because a difference in richness would require that new species appear or become extinct in removal plots over the course of a short growing season. In other words, removal treatments are likely to alter species richness over longer periods of time than in the time frame of a growing season. The effects of hemiparasitism by *Castilleja* on evenness, diversity and entropy suggest that the effects of this species alter the relative abundance of species in local communities, perhaps by altering competitive interactions among plants. So it seems possible that the presence of *Castilleja* promotes diversity and evenness, in much the same way that parasites, pathogens and predators in other systems promote diversity and evenness (e.g. Dobson and Crawley 1994).

The effects of hemiparasitism may depend on how selective *Castilleja* is for its host and how virulent it is in montane meadows and alpine ecosystems in general (Pennings and Callaway 2002). If *Castilleja* is largely a generalist, then it would parasitize multiple plant species and have little effect on specific plant species and plant community structure. For example, removing it should lead to a shift in the abundance of all its host species equally. However, if
Castilleja is primarily a specialist, then it would have a greater effect on the abundance of only one or a few species. So removing it would release those host species from parasitism, thereby increasing their competitive ability, and in turn causing a shift in the evenness, diversity, and entropy. In general, plots that had Castilleja present had higher species diversity measures except richness, so this study suggests that Castilleja may be more of a specialist hemiparasite than a generalist in alpine ecosystems. This makes logical sense, as many hemiparasites can selectively “forage” and “choose” which plants to parasitize. But what is surprising is that I did not detect any significant and consistent changes in the relative abundances of particular plant species resulting from Castilleja removal. Instead, what is likely happening is that within each plot, Castilleja specializes on a single or small number of species, but the identity and/or abundance of that species varied among plots.

The defining characteristic of hemiparasitic plants is that they have the ability both to make their own photosynthate and to parasitize their hosts (Press and Phoenix 2005). This means that they compete with other plants for light, water and nutrients and directly affect them through parasitism, so removing hemiparasites not only reduces their effect on other plants through parasitism, but it also reduces their competitive effect on their hosts. I tested for this competitive effect, however, using the random biomass removal treatment. If effects seen by the removal of hemiparasites were purely due to the reduction of competition with other plants, then the random biomass removal plots would react in the same ways as the Castilleja removal plots. Evenness, however, saw the most definitive changes due to the removal of Castilleja because the control and random biomass removal treatments were statistically the same, while Castilleja removal was significantly lower (Figure 1B). This confirms that the presence of Castilleja has effects on the rest of the plant community because of its hemiparasitic nature.
Parasitic and hemiparasitic plants are ubiquitous but have been largely excluded from community-level studies. Hemiparasites have the potential to influence community structure and composition, so it is important that plant community ecologists consider their roles in various ecosystems in order to obtain a better understanding of their potential impacts. Through several studies, it is understood that hemiparasites have varying effects on their plant communities depending on factors such as abundance or density of the hemiparasite, host preference, and soil fertility of the system. According to this study, the presence of Castilleja species generally increased the evenness, diversity, and entropy of the rest of the plant community. This is similar to the finding that Rhinanthus minor, also a hemiparasite, promoted plant diversity in a temperate grassland ecosystem (Bardgett et al. 2006). R. minor is a facultative root hemiparasite that, like Castilleja, affects plant community structure by promoting evenness (Bardgett et al. 2006). They found that plots with R. minor had 26% greater evenness (calculated using the inverse Simpson’s index) than those without the hemiparasite (Bardgett et al. 2006), and our study found that plots with Castilleja were 32% higher in evenness (calculated using the inverse Simpson’s index). The Bardgett study, however, was conducted in low-to-moderately productive grassland, so it is interesting to see how similar effects of hemiparasites can extend across different ecosystems. On the contrary, Spasojevic and Suding (2010) found that there was no significant difference in richness, evenness, and Simpson’s diversity index between plots with and without the hemiparasite Castilleja occidentalis in a dry meadow tundra.

The results of my study indicate that the hemiparasite Castilleja significantly increases evenness, diversity, and entropy in alpine ecosystems, enforcing the idea that hemiparasites can have detectable effects on plant community structure. Hemiparasites have largely been excluded from community-level studies, yet several studies show their importance in shaping and
changing communities (Bardgett et al. 2006; Press and Phoenix 2005; Spasojevic and Suding 2010). The influence of hemiparasites on their hosts may become more intense as global changes such as warming in these montane ecosystems increase, and understanding how hemiparasites regulate plant community composition may become increasingly important for making predictions on how ecosystems will function in the future.
ACKNOWLEDGEMENTS

This research was conducted as part of an undergraduate thesis project for Johannah Reed under the direction of Nathan Sanders at the University of Tennessee, Knoxville. I thank Nate Sanders for his constant guidance and expertise and for encouraging me to pursue ecology and become a part of his lab family. I also thank Aimee Classen and Lara Souza for making suggestions that greatly improved this research and paper. I extend my sincerest appreciation to Jennie Reithel, Tess Leuthner, Ross Brenan, and other students and faculty at the Rocky Mountain Biological Laboratory for their help with plant identification. I thank the Rocky Mountain Biological Laboratory for granting permission to stay and work at the lab. This research was funded by the Rocky Mountain Biological Laboratory, University of Tennessee Office of Research, Sanders Lab at the University of Tennessee, and the Chancellor’s Honors Program at the University of Tennessee.
LITERATURE CITED


**APPENDIX 1.** Abundance of all plant species in the plots based on data from the last observation at the end of the growing season. Percent (%) of plant species represented across all plots and for each treatment (Castilleja Removal, Control, Random Biomass Removal).

<table>
<thead>
<tr>
<th></th>
<th>Species</th>
<th>All</th>
<th><strong>Castilleja Removal</strong></th>
<th><strong>Control</strong></th>
<th><strong>Random Biomass Removal</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Poaceae</td>
<td>20.2</td>
<td>6.11</td>
<td>5.56</td>
<td>8.61</td>
</tr>
<tr>
<td>2</td>
<td>Paxistima myrsinites</td>
<td>11.6</td>
<td>5.28</td>
<td>2.50</td>
<td>3.89</td>
</tr>
<tr>
<td>3</td>
<td>Populus tremuloides</td>
<td>10.2</td>
<td>3.61</td>
<td>2.78</td>
<td>3.89</td>
</tr>
<tr>
<td>4</td>
<td>Castilleja miniata</td>
<td>9.50</td>
<td>0.00</td>
<td>5.56</td>
<td>3.94</td>
</tr>
<tr>
<td>5</td>
<td>Heterotheca villosa</td>
<td>6.28</td>
<td>2.28</td>
<td>3.39</td>
<td>0.61</td>
</tr>
<tr>
<td>6</td>
<td>Erigeron Speciosus</td>
<td>4.33</td>
<td>1.44</td>
<td>0.67</td>
<td>2.22</td>
</tr>
<tr>
<td>7</td>
<td>Potentilla hippiana</td>
<td>3.94</td>
<td>1.22</td>
<td>2.06</td>
<td>0.67</td>
</tr>
<tr>
<td>8</td>
<td>Helianthella quinqueneruis</td>
<td>3.94</td>
<td>1.17</td>
<td>2.78</td>
<td>0.00</td>
</tr>
<tr>
<td>9</td>
<td>Arenaria congesta</td>
<td>3.06</td>
<td>1.72</td>
<td>0.44</td>
<td>0.89</td>
</tr>
<tr>
<td>10</td>
<td>Mahonia repens</td>
<td>2.78</td>
<td>1.39</td>
<td>0.56</td>
<td>0.83</td>
</tr>
<tr>
<td>11</td>
<td>Lomatium dissectum</td>
<td>2.61</td>
<td>1.17</td>
<td>0.83</td>
<td>0.61</td>
</tr>
<tr>
<td>12</td>
<td>Vaccinium myrtillus</td>
<td>2.50</td>
<td>2.50</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td></td>
<td>Species</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
</tr>
<tr>
<td>---</td>
<td>-----------------------</td>
<td>----</td>
<td>----</td>
<td>----</td>
<td>----</td>
</tr>
<tr>
<td>1</td>
<td><em>Vicia americana</em></td>
<td>2.28</td>
<td>0.89</td>
<td>0.83</td>
<td>0.56</td>
</tr>
<tr>
<td>1</td>
<td><em>Viola praemorsa</em></td>
<td>2.00</td>
<td>0.67</td>
<td>0.72</td>
<td>0.61</td>
</tr>
<tr>
<td>1</td>
<td><em>Phacelia hastata</em></td>
<td>1.78</td>
<td>0.11</td>
<td>1.11</td>
<td>0.56</td>
</tr>
<tr>
<td>1</td>
<td><em>Artemisia tripartita</em></td>
<td>1.78</td>
<td>0.06</td>
<td>1.72</td>
<td>0.00</td>
</tr>
<tr>
<td>1</td>
<td><em>Senecio canus</em></td>
<td>1.39</td>
<td>0.17</td>
<td>0.83</td>
<td>0.39</td>
</tr>
<tr>
<td>1</td>
<td><em>Achillea millefolium</em></td>
<td>1.28</td>
<td>0.28</td>
<td>0.28</td>
<td>0.72</td>
</tr>
<tr>
<td>1</td>
<td><em>Lathyrus lanszwertii</em></td>
<td>1.11</td>
<td>0.11</td>
<td>0.56</td>
<td>0.44</td>
</tr>
<tr>
<td>2</td>
<td><em>Poa partensis</em></td>
<td>0.78</td>
<td>0.50</td>
<td>0.11</td>
<td>0.17</td>
</tr>
<tr>
<td>2</td>
<td><em>Galium Boreale</em></td>
<td>0.78</td>
<td>0.06</td>
<td>0.33</td>
<td>0.39</td>
</tr>
<tr>
<td>2</td>
<td><em>Draba aurea</em></td>
<td>0.61</td>
<td>0.56</td>
<td>0.00</td>
<td>0.06</td>
</tr>
<tr>
<td></td>
<td>Plant Species</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>---</td>
<td>------------------------------</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Phleum pratense</em></td>
<td>0.56</td>
<td>0.11</td>
<td>0.33</td>
<td>0.11</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Gentiana parryi</em></td>
<td>0.56</td>
<td>0.28</td>
<td>0.00</td>
<td>0.28</td>
</tr>
<tr>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Cymopterus lemmmonii</em></td>
<td>0.56</td>
<td>0.28</td>
<td>0.28</td>
<td>0.00</td>
</tr>
<tr>
<td>5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Potentilla gracilis</em></td>
<td>0.44</td>
<td>0.00</td>
<td>0.06</td>
<td>0.39</td>
</tr>
<tr>
<td>6</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Unknown Grass</em></td>
<td>0.33</td>
<td>0.06</td>
<td>0.28</td>
<td>0.00</td>
</tr>
<tr>
<td>7</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Rosa woodsii</em></td>
<td>0.33</td>
<td>0.06</td>
<td>0.00</td>
<td>0.28</td>
</tr>
<tr>
<td>8</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td><em>Unknown Herbaceous Plant #2</em></td>
<td>0.28</td>
<td>0.06</td>
<td>0.17</td>
<td>0.06</td>
</tr>
<tr>
<td>9</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td><em>Taraxacum officianale</em></td>
<td>0.28</td>
<td>0.00</td>
<td>0.00</td>
<td>0.28</td>
</tr>
<tr>
<td>0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td><em>Ipomopsis aggregata</em></td>
<td>0.28</td>
<td>0.11</td>
<td>0.00</td>
<td>0.17</td>
</tr>
<tr>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td><em>Unknown Herbaceous Plant #1</em></td>
<td>0.22</td>
<td>0.11</td>
<td>0.11</td>
<td>0.00</td>
</tr>
<tr>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td><em>Sedum lanceolatum</em></td>
<td>0.17</td>
<td>0.11</td>
<td>0.06</td>
<td>0.00</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td><em>Hydrophyllum capitatum</em></td>
<td>0.17</td>
<td>0.06</td>
<td>0.00</td>
<td>0.11</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td><em>Triteleia grandiflora</em></td>
<td>0.06</td>
<td>0.06</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td><em>Stipa richardsonii</em></td>
<td>0.06</td>
<td>0.00</td>
<td>0.06</td>
<td>0.00</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td><em>Frageria virginiana</em></td>
<td>0.06</td>
<td>0.00</td>
<td>0.06</td>
<td>0.00</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td><em>Arnica cordifolia</em></td>
<td>0.06</td>
<td>0.00</td>
<td>0.06</td>
<td>0.00</td>
</tr>
</tbody>
</table>