

**Identifying Pathogens of Switchgrass and Investigating
Antimicrobial Activity of Switchgrass-Derived Extractives**

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ABSTRACT

Switchgrass is an increasingly important biofuel crop, but knowledge of switchgrass fungal pathogens is not extensive. The purpose of this research was to identify the fungal pathogens that decrease crop yield of switchgrass grown in Tennessee and to investigate a potential value-added by-product of the switchgrass biofuel conversion process. The specific objectives were 1) to identify and characterize prevalent fungal pathogens of switchgrass in Tennessee, 2) assess switchgrass seed produced in the United States for seedborne fungal pathogens, and 3) evaluate switchgrass extractives for antimicrobial activity against plant pathogens.

Diseased switchgrass samples were collected from several locations in East Tennessee. Tissue was surface-sterilized and plated on potato dextrose agar (PDA), or water agar. Emergent fungi were isolated and identified based on colony, physiological, and molecular characteristics. Pathogenicity was confirmed with Koch's postulates in growth chamber studies. Nine pathogenic species were identified, several of which had not been reported on switchgrass previously.

To assess presence of seedborne pathogens, seed of seven cultivars from 11 commercial seed production companies located in the United States were tested, including multiple entries of 'Alamo', 'Blackwell', 'Cave-in-Rock', and 'Kanlow'. Three hundred surface-sterilized seed per lot were plated on PDA. Rates of fungal infection among the 30 sampled seed lots varied from less than 1 % to 87%. The most frequently observed pathogens were *Bipolaris oryzae*,

Alternaria alternata, and *Fusarium graminearum/ pseudograminearum*. Additional species of *Bipolaris* and *Fusarium* were present with less frequency.

To test the switchgrass extractives for antimicrobial activity, six ethanol-soluble extractives treatments, extracted from switchgrass collected from three farms at two different plant ages, were tested in a 3×2 factorial design for activity against four bacterial plant pathogens. Significant differences were shown for the main effect of bacteria ($P=0.09$) and the interaction effect of farm $\times$ plant age ($P=0.04$). *Xanthomonas perforans* was more sensitive to the extractives treatments than *Pseudomonas syringae* pv. *tomato* and extractives from 112 day-old switchgrass from farm C04 inhibited bacterial growth more than the extractives from 112 day-old switchgrass from farms C19 and C33.

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CHAPTER 1

LITERATURE REVIEW

Introduction

Switchgrass (*Panicum virgatum* L.) is a perennial native grass currently being investigated for use in production of cellulosic ethanol. Biofuel (cellulosic ethanol) from switchgrass has a low production cost and results in 540% more renewable energy than nonrenewable energy consumed (Schmer et al. 2008). Additionally, estimated greenhouse gas emissions from switchgrass-based cellulosic ethanol are 94% lower than estimated greenhouse gas emissions from gasoline (Schmer et al. 2008). The ethanol is produced through a biochemical process that uses various enzymes to convert the switchgrass cellulose to ethanol. To maximize the efficiency of this process, the plant extractives can be removed to allow for optimum enzyme activity (N. Labbé, personal communication, Anderson and Akin 2007). Switchgrass extractives are high in phenolic compounds (such as *p*-coumaric, ferulic, and sinapic acids), which are associated with the induction of the systemic resistance response that plants exhibit in the presence of pathogens or other stresses (Chen et al. 2010).

Although switchgrass-derived cellulosic ethanol production is a main focus of the biofuels initiative in Tennessee, limited research has been done on plant pathogens that occur on switchgrass grown in the state. As expansive monocultures of switchgrass are developed for commercial production, disease pressure is likely to increase due to lack of plant biodiversity (Wolfe 2000, Etheridge et al. 2001).

Host: *Panicum virgatum*

Switchgrass (*Panicum virgatum* L.) is a perennial, warm-season (C₄) tall prairie grass that has been adopted as a crop in the last 50 years (Parrish and Fike 2005). Switchgrass can grow up to 3 meters in height in one growing season and possesses a diffuse panicle seedhead with two-flowered spikelets at the end of long branches (Bouton 2008). Most switchgrass genotypes are caespitose (grow in small, dense clumps) and develop short rhizomes that allow the plant to form a loose sod over time (Bouton 2008). Switchgrass is native to North America and is adapted to a wide geographical range, spanning from 20° to 60° north latitude and east of 100° west longitude to the Atlantic seaboard (Vogel 2004). Two predominant ecotypes have developed: lowlands and uplands (Brunken and Estes 1975, Porter 1966, Vogel 2004). Basic chromosome number for both ecotypes is 9, but the somatic chromosome number is usually tetraploid in lowland selections and octaploid in uplands (Bouton 2008). Soil acidity typically is not a factor in switchgrass growth, but increased water-holding capacity as related to soil texture is necessary for stand establishment (Parrish and Fike 2005).

Several studies have demonstrated the importance of vesicular-arbuscular mycorrhizae in nutrient uptake for switchgrass (Clark and Zeto 2000, Hetrick et al. 1988, Boerner 1992, Brejda et al. 1998, Wilson et al. 2001). Brejda et al. (1998) tested growth and nutrient uptake in response to natural versus sterilized rhizosphere soil conditions. Switchgrass grown in soil with rhizosphere fungi and

bacteria produced 15-fold greater overall biomass, showed 6-fold greater nitrogen recovery and 36-fold greater phosphorus recovery than plants grown in sterile soil (Brejda et al. 1998). In several reviews, it has been suggested that mycorrhizae may mediate plant responses to drought stress, nutrient deficiencies, toxic metals, and pathogen attack (Parrish and Fike 2005).

Pathogens

More than 75 fungal pathogens occur on switchgrass in the United States (Farr and Rossman 2011). Close to 150 fungal isolates have been identified on switchgrass, but pathogenicity of many of these has not been determined (Ghmire et al. 2011, Gravert and Munkvold 2003). Twenty-four species of plant-parasitic nematodes, including species of *Dorylaimida*, *Triplonchida*, and *Tylenchida* and five viruses (*Panicum mosaic virus*, *Barley yellow dwarf virus*, *Sugarcane mosaic virus*, *Wheat soil-borne mosaic virus*, and *Maize rayado fino virus*) have been reported on switchgrass in the United States (Agindotan et al. 2010, Cassida et al. 2005, Farr and Rossman 2011, Garrett et al. 2004, Mekete et al. 2011, Sill 1957). In Tennessee, prior to published results of the current study, only one fungal disease had been reported: rust caused by *Puccinia emaculata* (Zale et al. 2008). The lack of reports of pathogens in Tennessee does not indicate a lack of presence. In 2008, researchers at the University of Tennessee found *Tilletia pulcherrima* (the causal agent of bunt disease) on switchgrass seed that had been produced in Texas and distributed to growers in

Tennessee (Carris et al. 2008). Based on reports from other regions in the United States (Farr and Rossman 2011), genera of fungal pathogens that are likely to occur in Tennessee include *Alternaria*, *Bipolaris*, *Curvularia*, and *Fusarium*.

Alternaria species can be difficult to identify due to conidial plasticity and low genetic variability among species (Misaghi et al. 1978, Kusaba and Tsunge 1995, Pryor and Gilbertson 2000). *Alternaria* includes nearly 100 species of dematiaceous mitosporic fungi that occur worldwide and range from general saprophytes to specific plant pathogens of cereals, ornamentals, nuts, vegetables, and fruits, including citrus (Thomma 2003). Spore production can be induced in the laboratory by use of specific culture media and environmental conditions (Shahin and Shephard 1979). Differential media assays have been developed for some *Alternaria* spp. (Andersen et al. 2001). Molecular identity can be confirmed by analysis of the internal transcribed spacer (ITS) region and mitochondrial small subunit (SSU) ribosomal DNA (Kusaba and Tsunge 1995, Pryor and Gilbertson 2000), or by random amplified polymorphic DNA (RAPD) analysis (Pryor and Gilbertson 2002, Roberts et al. 2000). Some anamorphic *Alternaria* species are associated with the teleomorphic genus *Lewia*, which is a member of the family Pleosporaceae in the phylum Ascomycota (Kirk 2011).

The genus *Bipolaris* (teleomorph: *Cochliobolus*) includes over 45 species that range from economically important pathogens of monocotyledonous hosts, such as wheat, barley, rice, and corn, to opportunistic human pathogens (Choudhry et al. 2010). On switchgrass, *B. oryzae*, *B. sorokiniana*, and *B.*

spicifera have been described as pathogenic in the United States (Farr and Rossman 2011, Krupinsky et al. 2004). Various species of *Bipolaris* are known to be seedborne and in 2004, *B. spicifera* was reported on grass seed exported from the United States to Korea (Koo et al. 2004).

Species of the genus *Fusarium* (teleomorph: *Gibberella* spp., *Nectria* spp., *Calonectria* spp., and *Micronectria* spp.) are a diverse array of mitosporic fungi, many of which are phytopathogenic to a wide range of plants under different environmental conditions (Booth 1971, Doohan et al. 2003). *Fusarium* species cause economically important diseases of monocotyledonous hosts such as *Fusarium* head blight of wheat and ear rot of corn (Parry et al. 1995, Sutton 1982). Some nonpathogenic *Fusarium* species are endophytic and can be added to soil to protect plants against fungal pathogens, including pathogenic species of *Fusarium* (Kavroulakis et al. 2007). *Fusarium* species have also been shown to have activity in *Fusarium* wilt-suppressive soils (Weller et al. 2002).

The genus *Curvularia* (teleomorph: *Cochliobolus*) includes species pathogenic to a wide range of hosts including wheat, rice, yam, mango, citrus, coconut and sorghum (de Luna et al. 2002). *Curvularia* spp. can be seedborne or soilborne and can cause primary infections, secondary infections, and post-harvest diseases (Lockwood 1988, Meehan 1947, Ray and Raavi 2005). *Curvularia lunata* has been found on grass seed exported from the United States to Korea (Koo et al. 2004) and *C. geniculata* has been identified causing secondary leaf spot on switchgrass in Kansas and Nebraska (Anonymous 1960).

In addition to their potential for decreasing crop yields, pathogens of switchgrass also are being investigated for use in the degradation of plant material in the conversion process of plant biomass to ethanol (Gibson et al. 2011). Plant pathogens may be preadapted for this new use as they have evolved numerous ways to degrade plant cell walls. Research is currently being conducted to exploit these mechanisms and enzymes for use in lignocellulose digestion of second-generation biofuels crops such as switchgrass.

Extractives

During the conversion process from cellulose to ethanol, the switchgrass biomass undergoes a series of biochemical reactions during which the switchgrass extractives are inhibitory to the process (Thammasouk et al. 1997). The ethanol-soluble switchgrass extractives contain phenolic compounds such as *p*-coumaric, ferulic, and sinapic acids, which are associated with antimicrobial activity, antioxidant activity, UV protection, and the induction of the systemic resistance response that plants exhibit in the presence of pathogens (Chen et al. 2010, Graf 1992, Herald and Davidson 1983, Kikuzaki et al. 2002, Nicholson and Hammerschmidt 1992, Walker 1994). In humans and mice, these phenolic compounds have also been shown to have anti-inflammatory activity, cholesterol-lowering capacity, and the ability to boost natural immune defenses (Chawla et al. 1987, Hu et al. 1990, Liu 1987).

Phenolic compounds can prevent plant diseases by inducing plant defense responses or by preventing pathogen growth (Nicholson and Hammerschmidt 1992). The responses of plants to pathogens have been differentiated based on host and non-host interactions, both of which are characterized by the early accumulation of phenolic compounds at the infection site (Fernandez and Heath 1989, Heath 1980). Commercial products are available that use plant extractives to induce resistance against both bacterial and fungal plant pathogens (Marrone BioInnovations 2009, Randoux et al. 2006). These products can be used in both conventional and organic crop production. For instance, Randoux et al. (2006) used giant-knotweed extract to inhibit fungal growth and enhance plant defenses in wheat inoculated with *Blumeria graminis* f. sp. *tritici* (Randoux et al 2006).

Research Objectives

The specific objectives of this study were: 1) to identify fungal pathogens that occur naturally on seedlings and mature switchgrass plants grown in Tennessee; 2) to identify seedborne pathogens of switchgrass; 3) to determine the antimicrobial potential of switchgrass extractives.

References

- Agindotan, B.O., Ahonsi, M.O., Gray, M.E., Bradley, C.A. 2010. Potential viral threats to *Miscanthus* × *giganteus* and switchgrass production for bioenergy in the United States. *Phytopathology* 100:S3.
- Andersen, B., Kroger, E., and Roberts, R.G. 2001. Chemical and morphological segregation of *Alternaria alternata*, *A. gaisen* and *A. longipes*. *Mycol. Res.* 105:291-299.
- Anderson, W.F., and Akin, D.E. 2007. Structural and chemical properties of grass lignocelluloses related to conversion for biofuels. *J. Ind. Microbiol. Biotechnol.* 35:355-366.
- Anonymous. 1960. Index of Plant Diseases in the United States. U.S.D.A. Agric. Handb. 165:1-531.
- Boerner, R.E. 1992. Plant life span and response to inoculation with vesicular-arbuscular mycorrhizal fungi. I. Annual versus perennial grasses. *Mycorrhiza* 1:153-161.
- Booth, C. 1971. The genus *Fusarium*. Commonwealth Mycological Institute. Kew, Surrey, England.
- Bouton, J. 2008. Improvement of switchgrass as a bioenergy crop. *Gen. Improv. Bioenerg. Crops* 11:296-308.
- Brejda, J.J., Moser, L.E., and Vogel, K.P. 1998. Evaluation of switchgrass rhizosphere microflora for enhancing seedling yield and nutrient uptake. *Agron. J.* 90:753-758.

- Brunken, J.N., and Estes, J.R. 1975. Cytological and morphological variation in *Panicum virgatum* L. Southwestern Naturalist 19:379-385.
- Carris, L.M., Castlebury, L.A., and Zale, J. 2008. First report of *Tilletia pulcherrima* on switchgrass (*Panicum virgatum* L.) in Texas. Plant Dis. 92:1707.
- Cassida, K.A. Kirkpatrick, T.L., Robbins, R.T., Muir, J.P., Venuto, B.C., and Hussey, M.A. 2005. Plant-parasitic nematodes associated with switchgrass (*Panicum virgatum* L.) grown for biofuel in the south central United States. Nematropica 35:1-10.
- Chawla, A.S., Singh, M., Murthy, M.S., Gupta, M.P., and Singh, H. 1987. Anti-inflammatory action of ferulic acid and its esters in carrageenan-induced rat paw edema model. Indian J. Exp. Biol. 25:187-189.
- Chen, S.F., Mowery, R.A., Sevcik, R.S., Scarlata, C.J., and Chambliss, C.K. 2010. Compositional analysis of water-soluble materials in switchgrass. J. Agric. Food Chem. 58:3251-3258.
- Choudhry, A., Tucci, V., and Greene, J.N. 2010. Disseminated Bipolaris infection. Infect. Dis. Clin. Pract. 19:204-207.
- Clark, R.B., and Zeto, S.K. 2000. Mineral acquisition by arbuscular mycorrhizal plants. J. Plant Nutrition 23:867-902.
- Clay, K. 1988. Fungal endophytes of grasses: a defensive mutualism between plants and fungi. Ecology 69:10-16.

- de Luna, L.Z., Watson, A.K., and Paulitz, T.C. 2002. Reaction of rice (*Oryza sativa*) cultivars to penetration and infection by *Curvularia tuberculata* and *C. oryzae*. Plant Dis. 86:470-476.
- Doohan, F.M., Brennan, J., and Cooke, B.M. 2003. Influence of climatic factors on *Fusarium* species pathogenic to cereals. Eur. J. Pl. Pathol. 109:755-768.
- Etheridge, J.V., Davey, L., and Christian, D.G. 2001. First report of *Rhizoctonia cerealis* causing sharp eyespot in *Panicum virgatum* in the UK. New Dis. Rep. 3:17.
- Farr, D.F., and Rossman, A.Y. Fungal Databases, Systematic Mycology and Microbiology Laboratory ARS, USDA. Retrieved October 28, 2011.
<<http://nt.ars-grin.gov/fungaldatabases/>>
- Fernandez, M.R. and Heath, M.C. 1989. Interaction of the non-host French bean plant (*Phaseolus vulgaris*) with parasitic and saprophytic fungi. III. Cytologically detectable responses. Can. J. Bot. 67:676-686.
- Garrett, K.A., Dendy, S.P., Power, A.G., Blaisdell, G.K., Alexander, H.M., and McCarron, J.K. 2004. Barley yellow dwarf disease in natural populations of dominant tallgrass prairie species in Kansas. Plant Dis. 88:574.
- Ghimire, S.R., Charlton, N.D., Bell, J.D., Krishnamurthy, Y.L., and Craven, K.D. 2011. Biodiversity of fungal endophyte communities inhabiting switchgrass (*Panicum virgatum* L.) growing in native tallgrass prairie of northern Oklahoma. Fung. Diversity 47:19-27.

- Gibson, D.M., King, B.C., Hayes, M.L., and Bergstrom, G.C. 2011. Plant pathogens as a source of diverse enzymes for lignocellulose digestion. *Curr. Opin. Microbiol.* 14:264-270.
- Graf, E. 1992. Antioxidant potential of ferulic acid. *Free Radical Biol. Med.* 13:435-448.
- Gravert, C.E. and Munkvold, G.P. 2002. Fungi and diseases associated with cultivated switchgrass in Iowa. *J. Iowa Acad. Sci.* 109: 30-34.
- Heath, M.C. 1980. Reactions of nonsusceptible to fungal pathogens. *Annu. Rev. Phytopathol.* 18:211-36
- Herald, P.J., and Davidson, P.M. 1983. Antibacterial activity of selected hydroxycinnamic acids. *J. Food Sci.* 48:1378–1379.
- Hetrick, B.A., Kitt, D.G., and Wilson, G.T. 1988. Mycorrhizal dependence and growth habit of warm-season and cool-season tallgrass prairie plants. *Can. J. Botany.* 66:1376-1380.
- Hu, H., Hang, B., and Wang, P. 1990. Anti-inflammatory effects of ferulic acid. *Zhongguo Yaoke Daxue Xuebao* 21:279-282.
- Kavroulakis, N., Ntougias, S., Zervakis, G.I., Ehalotis, C., Haralampidis, K., and Papadopoulou, K.K. 2007. Role of ethylene in the protection of tomato plants against soil-borne fungal pathogens conferred by an endophytic *Fusarium solani* strain. *J. Exp. Bot.* 58:3853-3864.

- Kikuzaki, H., Hisamoto, M., Hirose, K., Akiyama, K., and Taniguchi, H. 2002. Antioxidant properties of ferulic acid and its related compounds. J. Agric. Food Chem. 50:2161-2168.
- Kirk, P. 2011. Index Fungorum. CABI, retrieved October 31, 2011.
- Koo, H.M., Lee, S.H., Chung, I.M., and Chun, S.E. 2004. Cultural characteristics of a seedborne fungus, *Bipolaris spicifera* detected from imported grass seeds into Korea. Mycobiology 32:186-189.
- Krupinsky, J.M., Berdahl, J.D., Schoch, C.L., and Rossman, A.Y. 2004. Leaf spot on switchgrass (*Panicum virgatum*), symptoms of a new disease caused by *Bipolaris oryzae*. Can. J. Plant Pathol. 26:371-378.
- Kusaba, M., and Tsuge, T. 1995. Phylogeny of *Alternaria* fungi known to produce host-specific toxins on the basis of variation in internal transcribed spacers of ribosomal DNA. Curr. Genet. 28:491-498.
- Lockwood, J.L. 1988. Evolution of concepts associated with soilborne plant pathogens. Annu. Rev. Phytopathol. 26:93-121.
- Liu, Y. 1987. Pharmaceutical composition for increasing immunity and decreasing side effects of anticancer chemotherapy. U.S. Patent 4687761.
- Marrone BioInnovations. 2009. Retrieved October 31, 2011. <<http://www.marronebioinnovations.com/products/regalia/>>.
- Meehan, F. 1947. A host index to seed-borne species of *Helminthosporium* and *Curvularia* on certain grasses. Proc. Assoc. Offic. Seed Analysts 37:89-92.

- Mekete, T., Reynolds, K., Lopez-Nicora, H. D., Gray, M. E., and Niblack, T. L. 2011. Plant-parasitic nematodes are potential pathogens of *Miscanthus × giganteus* and *Panicum virgatum* used for biofuels. *Plant Dis.* 95:413-418.
- Misaghi, I.J., Grogan, R.G., Duniway, J.M., and Kimble, K.A. 1978. Influence of environment and culture media on spore morphology of *Alternaria alternata*. *Phytopathology* 68:29-34.
- Nicholson, R.L., and Hammerschmidt, R. 1992. Phenolic compounds and their role in disease resistance. *Annu. Rev. Phytopathol.* 30:369-389.
- Parrish, D.J., and Fike, J.H. 2005. The biology and agronomy of switchgrass for biofuels. *Crit. Rev. Plant Sci.* 24:423-459.
- Parry, D.W., Pettitt, T.R., Jenkinson, P., and Lees, A.K. 1995. The cereal *Fusarium* complex. Pages 301-320 in: *Ecology of Plant Pathogens*, P. Blakeman and B. Williamson, eds. CAB International, Wallingford, UK.
- Porter, C.L. 1966. An analysis of variation between upland and lowland switchgrass *Panicum virgatum* L. in central Oklahoma. *Ecology* 47:980-992.
- Pryor, B.M., and Gilbertson, R.L. 2000. Molecular phylogenetic relationships amongst *Alternaria* species and related fungi based upon analysis of nuclear ITS and mt SSU rDNA sequences. *Mycol. Res.* 104:1312-1321.
- Pryor, B.M., and Gilbertson, R.L. 2002. Relationships and taxonomic status of *Alternaria radicina*, *A. carotiincultae*, and *A. petroselini* based upon

- morphological, biochemical, and molecular characteristics. *Mycologia* 94:49-61.
- Randoux, B., Renard, D., Nowak, E., Sanssene, J., Courtois, J., Durand, R. and Reignault, P. 2006. Inhibition of *Blumeria graminis* f. sp. *tritici* germination and partial enhancement of wheat defenses by Milsana. *Phytopathology* 96:1278-1286.
- Ray, R.C., and Raavi, V. 2005. Post-harvest spoilage of sweet potato in tropics and control measures. *Crit. Rev. Food Sci. Nutr.* 45:623-644.
- Roberts, R.G., Reymond, S.T., and Andersen, B. 2000. RAPD fragment pattern analysis and morphological segregation of small-spored *Alternaria* species and species groups. *Mycol. Res.* 104:151-160.
- Schmer, M.R., Vogel, K.P., Mitchell, R.B., and Perrin, R.K. 2008. Net energy of cellulosic ethanol from switchgrass. *PNAS* 105:464-469.
- Shahin, E.A., and Shephard, J.F. 1979. An efficient technique for inducing profuse sporulation of *Alternaria* species. *Phytopathology* 69:618-620.
- Sill, W.H. 1957. Panicum mosaic, a new virus disease of *Panicum virgatum* and related grasses. *Phytopathology* 47:31.
- Sutton, J.C. 1982. Epidemiology of wheat head blight and maize ear rot caused by *Fusarium graminearum*. *Can. J. Plant Pathol.* 4:195–209.
- Thammasouk, K., Tandjo, D., and Penner, M.H. 1997. Influence of extractives on the analysis of herbaceous biomass. *J. Agric. Food Chem.* 45:437-443.

- Thomma, B.P. 2003. *Alternaria* spp.: from general saprophyte to specific parasite. Molec. Plant Pathol. 4:225-236.
- Vogel, K.P. 2004. Switchgrass. Pages 561-588 in: Warm-season (C4) grasses. Agron. Monogr. 45. ASA, CSSA, and SSSA, L.E. Moser, B.L. Burson and L.E. Sollenberger, eds. Madison, WI.
- Walker, J.R. 1994. Antimicrobial compounds in food plants. Pages 181-204 in: Natural Antimicrobial Systems and Food Preservation. V.M. Dillon and R.G. Board, eds., CAB International, Wallingford, UK.
- Welbaum, G.E., Sturz, A.V., Dong, Z., and Nowak, J. 2004. Managing soil microorganisms to improve productivity of agro-ecosystems. Crit. Rev. Plant Sci. 23:175-193.
- Weller, D.M., Raaijmakers, J.M., McSpadden-Gardener, B.B., Thomashow, L.S. 2002. Microbial populations responsible for specific soil suppressiveness to plant pathogens. Annu. Rev. Phytopathol. 40:309-48.
- Wolfe, M.S. 2000. Crop strength through diversity. Nature 406:681-682.
- Zale, J., Freshour, L., Agarwal, S., Sorochan, J., Ownley, B.H., Gwinn, K.D., Castlebury, L.A., and Carris, L.M. 2008. First report of *Puccinia emaculata* on switchgrass (*Panicum virgatum* L.) in Tennessee. Plant Dis. 92:1710.

CHAPTER 2

IDENTIFICATION OF FUNGAL PATHOGENS OF AGRONOMIC SWITCHGRASS GROWN IN TENNESSEE

Parts of this chapter were adapted from the following publications:

- Vu, A.L., Dee, M.M., Gualandi, Jr., R.J., Huff, S., Zale, J., Gwinn, K.D., and Ownley, B.H. 2011. First report of leaf spot caused by *Bipolaris spicifera* on switchgrass in the United States. Plant Dis. 95:1191.
- Vu, A.L., Gwinn, K.D., and Ownley, B.H. 2011. First report of spot blotch and common root rot caused by *Bipolaris sorokiniana* on switchgrass in Tennessee. Plant Dis. 95:1195.
- Vu, A.L., Gwinn, K.D., and Ownley, B.H. 2011. First report of *Sclerotinia homoeocarpa* causing dollar spot on switchgrass in the United States. Plant Dis. 95:1585.
- Vu, A.L., Dee, M.D., Russell, T., Zale, J., Gwinn, K.D., and Ownley, B.H. First report of leaf spot caused by *Alternaria alternata* on switchgrass in Tennessee. Plant Dis. (Accepted).

M.M. Dee, R.J. Gualandi, Jr., and S. Huff assisted in fungal collection, isolation and morphological identification of *Bipolaris spicifera*, and determination of pathogenicity. M.M. Dee and T. Russell assisted in collection and identification of *Alternaria alternata*. J. Zale, K.D. Gwinn, and B.H. Ownley provided guidance and funding for the studies.

Abstract

Switchgrass is a perennial warm-season native grass currently being investigated for use in biomass-based ethanol production in Tennessee. However, little is known about fungal diseases that occur, or the impact of these diseases on crop yield. Disease reduction could significantly increase biomass yield and overall crop quality, particularly in the southeastern U.S. where large monocultures are being planted. The goal of this project was to identify fungal pathogens of switchgrass to form a foundation for understanding of role of disease in the overall efficiency and sustainability of switchgrass as a biofuel crop. Naturally infected 'Alamo' switchgrass plants were collected from growers' fields in Vonore, TN in summer 2009 and agronomic research plots and ornamental plantings in Knoxville, TN in winter 2007 through spring 2008, and again in summer 2010. Fungi were isolated from diseased plants and pathogenicity was confirmed with Koch's postulates in growth chamber studies with whole plant assays. Pathogenic species of *Alternaria*, *Bipolaris*, and *Fusarium* were identified; several of these species had not been described previously on switchgrass, but are known to reduce quality and yield of other monocotyledonous crops. Species identification was based on morphology of conidia and conidiogenous cells, colony characteristics, colony growth at various temperatures, and confirmed by comparison of internal transcribed spacer (ITS) and mitochondrial small subunit (SSU) sequences of ribosomal DNA with

nucleotide sequences in the National Center for Biotechnology Information (NCBI) GenBank database.

Introduction

Switchgrass (*Panicum virgatum* L.) is a perennial grass currently being investigated for use as a biomass feedstock for cellulosic ethanol production. Switchgrass biofuel has a low production cost and yields much more renewable energy than nonrenewable energy consumed in the process of production. Also, estimated greenhouse gas emissions from switchgrass-derived ethanol are 94% lower than estimated greenhouse gas emissions from gasoline (Schmer et al. 2008).

Although switchgrass-derived cellulosic ethanol production is a main focus of the biofuels initiative in Tennessee, only limited research has been done on pathogens that occur on switchgrass grown in the state. As expansive monocultures of switchgrass are developed for commercial production, disease pressure is likely to increase due to lack of plant biodiversity in those fields.

Previous studies have shown that nearly 150 species of fungi occur on switchgrass; with 75 confirmed pathogens in the United States (Farr and Rossman 2011). Prior to this study, only one fungal disease had been reported in Tennessee: rust caused by *Puccinia emaculata* (Zale et al. 2008). The lack of reports of switchgrass pathogens in Tennessee does not indicate a lack of

presence. In 2008, researchers at the University of Tennessee found *Tilletia pulcherrima* (causal agent of bunt disease) on switchgrass seed that had been produced in Texas and distributed to growers in Tennessee (Carris et al. 2008). The purpose of this research was to develop information on fungal pathogens that decrease crop yield and quality of switchgrass grown in Tennessee.

Materials and Methods

Mature 'Alamo' switchgrass plants, with symptoms of leaf, stem, or root disease, were collected from agronomic research plots or ornamental plantings in Knoxville, TN in winter 2007 through spring 2008, and again in summer 2010. Naturally-infected switchgrass seedlings and mature plants also were collected from agronomic stands in Vonore, TN, in summer 2009. Plant tissue was surface-sterilized with 95% ethanol for 1 min, 20% commercial bleach (NaClO₃; Clorox, Oakland, CA) for 2 min, 95% ethanol for 1 min, dried on sterile filter paper, and plated on 2% water agar amended with 10 mg/liter rifampicin (Sigma-Aldrich, St. Louis, MO) and 5 to 10 µl/liter miticide (2.4 EC Danitol, Valent Chemical, Walnut Creek, CA). Fungal hyphae were transferred to potato dextrose agar (PDA) for axenic culture and identified based on morphology of conidia, conidiogenous cells, and colony growth. In some cases, isolates were grown on specialized culture media and were cultured at different incubation temperatures to determine identification.

Some fungi (*Alternaria alternata*, *Bipolaris sorokiniana*, *Pithomyces chartarum*) were isolated from plants grown in growth chambers intended for pathogenicity studies, indicating that the isolated pathogen had been seedborne. 'Alamo' seed (Seedland, Wellborn, FL) had been surface-sterilized with 70% Clorox for 30 min, washed with sterile water, and dried prior to planting. Organisms were isolated and identified as described for pathogens from diseased tissues. All seed planted for subsequent pathogenicity tests were scarified with 60% sulfuric acid then surface-sterilized with 50% Clorox for 30 min, washed with sterile water, and shaken in sterile water for 1 to 3 days prior to planting (Gwinn et al. 1991).

For pathogenicity assays, seed were sown in 9×9-cm pots containing 50% ProMix Potting and Seeding Mix (Premier Tech Horticulture, Rivière-du-Loup, Québec, Canada) and 50% Turface ProLeague (Profile Products, Buffalo Grove, IL) (vol/vol). A conidial spore suspension (10^4 to 10^7 spores/ml water) or mycelial slurry was made from each fungus and sprayed on 6 to 10 replicates of switchgrass seedlings in a growth chamber. Plants were kept at high humidity for 2 to 7 days at 25/20°C with a 12-hour photoperiod. Symptomatic plant tissue was collected. Fungi were re-isolated and identified as described above to fulfill Koch's postulates.

Genomic DNA was extracted from each isolate and re-isolate (obtained from the Koch's postulates pathogenicity assay) with the DNeasy Plant Mini Kit (Qiagen, Valencia, CA). Ribosomal DNA from the internal transcribed spacer

(ITS) region was amplified with polymerase chain reaction (PCR) with primers ITS4 and ITS5 (White et al. 1990). The DNA fragments were sequenced and checked against the GenBank database with BLASTn (NCBI). For cases in which the ITS sequence was not specific enough for an exact GenBank match, DNA from an internal region of the mitochondrial small subunit (SSU) gene was amplified with PCR with primers NMS1 and NMS2 (Li et al. 1994). DNA fragments were sequenced and checked against the GenBank database with BLASTn (NCBI).

***Alternaria alternata* (Fries) Keissler**

‘Alamo’ switchgrass grown in Vonore, TN was found exhibiting light brown-to-dark brown spots and general chlorosis in June, 2009. Symptomatic leaf tissue samples were surface-sterilized and plated on 2% water agar amended with antibiotic and miticide as described earlier. Plates were incubated at 26°C for 4 days in darkness. An asexual, dematiaceous mitosporic fungus was isolated and transferred to PDA. Cultures were transferred to *Alternaria* sporulation medium (Shahin and Shephard 1979) to induce conidial production and to dichloran rose bengal yeast extract sucrose agar to determine morphological features and colony growth characteristics (Andersen et al. 2001, Simmons 1999). Pathogenicity studies were conducted with nine pots of approximately 20 plants each of 5-week-old ‘Alamo’ switchgrass plants grown from surface-sterilized seed. Plants were wounded by trimming the tops to a

height of approximately 5 cm. Eight pots were sprayed with a conidial spore suspension of 5.07×10^6 spores/ml sterile water and subjected to high humidity by enclosure in a plastic bag for 7 days. One pot was sprayed with sterile water and subjected to the same conditions to serve as a control. Plants were maintained in a growth chamber at 25/20°C with a 12-h photoperiod. Lesions were excised, surface-sterilized, plated on water agar, and identified in the same manner as previously described. The ITS region of ribosomal DNA and the mitochondrial SSU region from the original isolate and the re-isolate recovered from the pathogenicity assay were amplified with PCR, with primers ITS4 and ITS5 and NMS1 and NMS2, respectively.

Bipolaris oryzae (Breda de Haan) Shoemaker

Dark brown to black irregular-shaped foliar spots were observed on 'Alamo' switchgrass grown on the campus of the University of Tennessee in December 2007. Symptomatic leaf tissue samples were surface-sterilized and plated on 2% water agar amended with antibiotic and miticide as described earlier. An asexual, sparsely-sporulating, dematiaceous mitosporic fungus was observed and fungal plugs were transferred to surface-sterilized detached leaves on sterile filter paper in a moist chamber to increase spore production. Disease assays were conducted with 6-week-old switchgrass plants grown from seed scarified with 60% sulfuric acid and surface-sterilized in 50% bleach (Gwinn et al. 1991). Nine pots with approximately 20 plants per pot were inoculated with a

mycelial slurry. Two additional pots were inoculated with sterile water and subjected to the same conditions to serve as a control. Plants were subjected to high humidity by enclosure in a plastic bag for 72 h. Bags were removed and plants were incubated at 25/20°C with 50 to 60% relative humidity. During the incubation period, plants were maintained in a growth chamber with a 12-h photoperiod of fluorescent and incandescent lighting. Symptomatic leaf tissue was processed and plated as described above. The ITS region of ribosomal DNA from the original isolate and from the isolate recovered from plants in the pathogenicity assay was amplified with PCR, with primer pairs ITS4 and ITS5.

***Bipolaris sorokiniana* (Saccardo) Shoemaker**

Light to dark brown, irregular-shaped leaf spots, chlorosis, necrotic roots, and severe stunting were observed on 'Alamo' switchgrass grown on the campus of the University of Tennessee in December 2007. Symptomatic leaf and root samples were surface-sterilized and plated onto water agar amended with antibiotic and miticide as described earlier. Plates were incubated at 25°C in darkness for 4 days. A sporulating, dematiaceous mitosporic fungus was noted and transferred to PDA. Disease assays were conducted with 5-week-old 'Alamo' switchgrass grown from surface-sterilized seed. Ten pots with approximately 20 seedlings were sprayed with 2.4×10^5 spores/ml of sterile water, with one control plant sprayed with sterile water and subjected to the same conditions as treated plants. Plants were subjected to high humidity created by enclosure in a plastic

bag for 45 h. The bag was removed and plants were incubated at 25/20°C with 50 to 60% relative humidity. During the incubation, plants were maintained in a growth chamber with a 12-h photoperiod of fluorescent and incandescent lighting. Foliar lesions and diseased roots were surface-sterilized, plated on water agar, and the resultant fungal colonies were processed as described earlier. The ITS and mitochondrial SSU regions of ribosomal DNA from the original isolate, and the isolate recovered from plants in the pathogenicity assay, were amplified with PCR, with primer pairs ITS4 and ITS5, and NMS1 and NMS2, respectively.

***Bipolaris spicifera* (Bainier) Subramanian**

Light to dark brown leaf spots and general chlorosis were observed on 'Alamo' switchgrass grown in ornamental plantings on the campus of the University of Tennessee in Knoxville in December 2007. Disease distribution was patchy, infecting approximately 10% of plants. Patches had mild to severely infected plants, with stunting in areas of severe infection. Symptomatic leaf tissue was surface-sterilized and plated onto water agar amended with antibiotic and miticide as described earlier. Plates were incubated at 26°C in darkness for 5 days. A sporulating, dematiaceous mitosporic fungus was observed and transferred to PDA. Pathogenicity studies were conducted with 5-week-old 'Alamo' switchgrass plants grown from surface-sterilized seed. Ten replicate pots with approximately 20 plants each were sprayed with a spore suspension of $4.5 \times$

10⁶ spores/ml of sterile water prepared from 6-day-old cultures grown on PDA. Plants were subjected to high humidity for 45 h by enclosure in a plastic bag, and then incubated at 25/20°C with a 12-h photoperiod in a growth chamber. Two control pots were inoculated with sterile water and subjected to the same conditions. Lesions were excised from leaves, surface-sterilized, plated on water agar for morphological identification. The ITS region of ribosomal DNA from the original isolate used for inoculation and the re-isolated culture recovered from plants in the pathogenicity studies were amplified with PCR using primers ITS4 and ITS5 (White 1990).

Fusarium acuminatum Ellis & Everhart

In June 2009, 'Alamo' switchgrass leaf and stem samples with brown leaf spots and streaks were collected from agronomic switchgrass plots in Vonore, Tennessee. Infected tissue was surface-sterilized and plated on 2% water agar amended antibiotic and miticide as described earlier. Plates were incubated for two days at 22.5 °C, and the fungus was transferred to PDA. Cultures were incubated at 22.5 °C in the dark for 5-7 days and on carnation leaf agar (CLA) for further identification. Morphological identification was enhanced by incubating cultures on PDA at 25°C with no light, 30°C with light, and alternating between 25° with light for 12 h and 20°C in darkness for 12 h. Pathogenicity was determined by inoculation of ten pots containing approximately 20 healthy switchgrass plants grown from surface-sterilized seeds. Leaves were wounded

by trimming the tops and sprayed with 1.1×10^6 spores/ml water with an atomizer. Conidial solution was also sprayed on the crown of the plants at the soil surface. Two untreated pots were sprayed with sterile water to serve as controls. Plants were subjected to high humidity by enclosure in a plastic bag. Bags were removed after 24 h and plants were incubated at 27°C in a growth chamber. The ITS region of ribosomal DNA and mitochondrial SSU DNA from the original isolate and the re-isolated culture were amplified with PCR using primers ITS4 and ITS5, and NMS1 and NMS2, respectively.

***Fusarium equiseti* (Corda) Saccardo**

Light brown to brown leaf spots, general chlorosis, and stunting were observed on 'Alamo' switchgrass grown on the campus of the University of Tennessee in December 2007. Infected root and crown tissue was surface-sterilized and plated on 2% water agar amended with antibiotic and miticide as described earlier. Plates were incubated for two days at 22.5°C and emergent fungal growth was transferred to PDA. Cultures were incubated in darkness at 22.5°C on PDA and on CLA for identification. Pathogenicity was determined by inoculation of nine pots containing approximately 20 healthy switchgrass plants grown from surface-sterilized seeds. Leaves were wounded by trimming the tops to a height of approximate 5 cm and roots were wounded with a sterile surgical blade, and then sprayed with 4.7×10^6 spores/ml water with an atomizer on and around the crown of the plant at the soil surface. Two control pots were sprayed

with sterile water to serve as controls. Plants were subjected to high humidity by enclosure in a plastic bag. Bags were removed after 48 h and plants were incubated at 27°C in a growth chamber. The pathogen was re-isolated from symptomatic leaf and root tissue and identified on PDA based on spore morphology as described above. The ITS region of ribosomal DNA from the original isolate and the re-isolate was amplified with PCR using primers ITS4 and ITS5.

***Fusarium pseudograminearum* Aoki & O'Donnell**

In June 2009, 'Alamo' switchgrass leaf and stem samples showing brown leaf spots and streaks were collected from agronomic switchgrass plots in Vonore, Tennessee. Infected stem tissue was surface-sterilized and plated on 2% water agar amended with antibiotic and miticide as described earlier. Plates were incubated for 2 days at 22.5°C. The resultant fungus was transferred to PDA and CLA for morphological identification.

Pathogenicity was determined by inoculation of eight pots containing approximately 20 healthy switchgrass plants grown from surface-sterilized seeds. Leaves were wounded by trimming the tops to a height of 5 cm, and then sprayed with 3.2×10^5 spores/ml water with an atomizer. Conidial solution was sprayed also on the crown of the plants at the soil surface. Two pots of plants were sprayed with sterile water to serve as controls. Plants were subjected to high humidity by enclosure in a plastic bag. Bags were removed after 48 h and

plants were incubated at 26°C in a growth chamber. The ITS region of ribosomal DNA from the original isolate and re-isolate recovered from plants in the pathogenicity studies were amplified with PCR using primers ITS4 and ITS5.

***Pithomyces chartarum* (Berkeley & M.A. Curtis) M.B. Ellis**

Light brown to white bleached spots were observed on 'Alamo' switchgrass grown from surface-disinfested seed grown in Colorado in fall 2009. Symptomatic leaf tissue was surface-sterilized, air-dried on sterile filter paper, and plated on 2% water agar amended with antibiotic and miticide as described earlier. Plates were incubated at 26°C in darkness for 5 days. A sporulating, dematiaceous mitosporic fungus was observed and transferred to PDA. Pathogenicity studies were conducted with 5-week-old 'Alamo' switchgrass plants grown from seed scarified with 60% sulfuric acid and surface-sterilized with 50% bleach. Eight replicate pots with approximately 20 plants each were sprayed with a spore suspension of 5.7×10^5 spores/ml of sterile water prepared from 6-day-old cultures grown on V8-juice agar in the dark. Plants were subjected to high humidity for 72 h by enclosure in a plastic bag, and then incubated at 25/20°C with a 12-h photoperiod in a growth chamber. Lesions were excised from leaves, surface-sterilized, plated on water agar, and the resulting cultures were grown on PDA for morphological identification. The ITS region of ribosomal DNA from the original isolate and re-isolate were amplified with PCR using primers ITS4 and ITS5.

***Sclerotinia homoeocarpa* F.T. Bennett**

Sclerotinia homoeocarpa causes dollar spot on many grass species; however it has not been described on switchgrass as a host. In August 2010, bleached, tan to straw-colored leaf spots with dark brown to reddish-brown margins were found in patchy distribution in small field plots of 'Alamo' switchgrass at the East Tennessee Research and Education Center, Knoxville, TN. The plots had been planted to switchgrass for the past 21 years. Disease lesions covered 75-80% of leaf tissue per patch and were also evident on stems. To identify the pathogen, center portions of diseased leaves were cut into 20- to 30-cm-long segments, and surface-disinfested, then 5-cm-long sections that included a leading edge of a lesion were plated on PDA. Plates were incubated at 22°C.

Pathogenicity studies were conducted with 6-week-old 'Alamo' switchgrass grown from scarified and surface-disinfested seed (Gwinn et al. 1991). Nine pots with 18 plants each were inoculated with 20 mycelial plugs (6-mm-diameter) per pot, taken from 3-to-5-day-old fungal cultures. Two control pots were inoculated with sterile PDA plugs and subjected to the same conditions. Plugs were placed on leaf surfaces and around plant crowns. Plants were subjected to high humidity by enclosure in a plastic bag and incubated in a growth chamber at 25/20°C with a 12-h photoperiod. Plastic bags were removed after 48 h. The fungus was cultured from leaf spots and stem lesions of inoculated plants as described above. The same disease and fungus were

observed, completing Koch's postulates. The ITS region of ribosomal DNA from the original isolate and re-isolate was amplified with PCR, with primers ITS4 and ITS5. The mitochondrial SSU was amplified from the original isolate with primers NMS1 and NMS2.

Results and Discussion

Alternaria alternata (Fries) Keissler

Club-shaped conidia were produced in chains, with branching of chains present on *Alternaria* sporulation medium (Fig. 1). Conidia were 27 to 50 μm in length $\times$ 10 to 15 μm in width (mean $42.5 \times 12.5 \mu\text{m}$). Morphological features and growth on dichloran rose bengal yeast extract sucrose agar were consistent with the characteristics described previously for *A. alternata* (Fr.) Keissl. (Andersen et al. 2001, Simmons 1999).

Foliar leaf spot symptoms appeared 5 to 10 days post-inoculation for all pots inoculated with *A. alternata* in the pathogenicity assay. Symptoms of *A. alternata* infection were not observed on the control. The ITS region sequences of the original isolate and the re-isolate were approximately 537 bp and were submitted to GenBank (Accession Nos. HQ130485.1 and HQ130486.1). The BLAST search (BLASTn, NCBI) against GenBank isolates indicated a match of 100% maximum identity with eight *A. alternata* isolates, including GenBank Accession No. AB470838. The SSU sequences were approximately 551 bp and

matched 100% maximum identity for seven *A. alternata* isolates, including GenBank Accession No. AF229648. *Alternaria alternata* has been reported from switchgrass in Iowa (Gravert and Munkvold 2002); however, pathogenicity was not confirmed with Koch's postulates in that report. To our knowledge, this is the first report of *A. alternata* causing leaf spot on switchgrass in Tennessee.

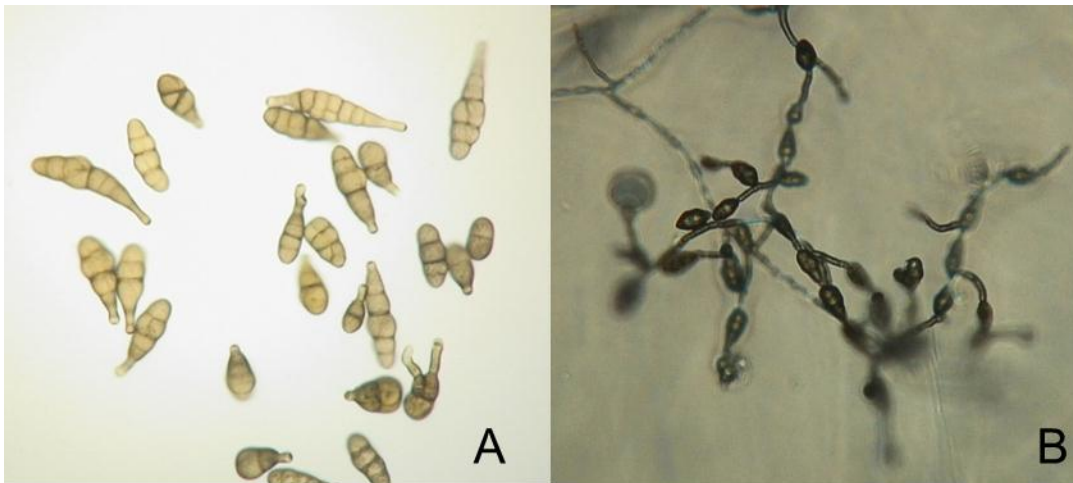


Fig. 1. A. Conidia of *Alternaria alternata*, B. Acropetal conidial ontogeny with branching of *A. alternata*.

***Bipolaris oryzae* (Breda de Haan) Shoemaker**

Conidia were ovate, oblong, mostly straight to slightly curved, and light to olive-brown with three to ten septa (Fig. 2). Conidial dimensions were 12.5×27.5 to 17×95 (average 14.5×72) μm . Conidia were produced on single, light brown, multiseptate conidiophores that were polytretic, geniculate, and sympodial. Morphological characteristics and disease symptoms were similar to those

described for *B. oryzae* (Breda de Haan) Shoemaker (Sivanesan 1987, Krupinsky et al. 2004).

In the pathogenicity assay, foliar leaf spot symptoms appeared 5 to 14 days post-inoculation for eight of nine replicates. Control plants showed no symptoms. PCR amplicons of the ITS region were approximately 534 bp. Amplicon sequences from the original isolate and re-isolate were identical and had 99% homology to several *B. oryzae* isolates from switchgrass in Mississippi (GenBank Accession Nos. GU222690, GU222691, GU222692). Leaf spot caused by *B. oryzae* on switchgrass has been described in North Dakota, Mississippi, and New York (Farr and Rossman 2011) and the fungus has been shown to have seedborne transmission (Tomaso-Peterson and Balbalian 2010).



Fig. 2. Conidia of *Bipolaris oryzae*.

***Bipolaris sorokiniana* (Saccardo) Shoemaker**

On PDA, conidia were ovate, oblong, mostly straight, and olive to brown with three to nine septa (Fig. 3). Conidial dimensions were 12.5×27.5 (17.5) to 20×77.5 (57) μm . Conidia were produced on single, light brown, multiseptate conidiophores that were polytretic, geniculate, and sympodial. Morphological features were as described for *B. sorokiniana* (Sacc.) Shoemaker (teleomorph = *Cochliobolus sativus*) (Nyvall and Percich 1999, Sivanesan and Holliday 1981).

In the pathogenicity assay, foliar leaf spot symptoms appeared 6 to 10 days post-inoculation for plants in all 10 replicates and necrotic lesions were observed on roots. Resultant colonies were identified as *B. sorokiniana*.

PCR amplicons of the ITS and SSU regions of approximately 551 and 571 bp were obtained, respectively. Both amplicons were obtained from each isolate and sequenced. Amplicon sequences from the original isolate and re-isolate were identical and the sequences were submitted to GenBank (Accession Nos. HQ611957 and HQ611958). The ITS sequences had 98% homology to 23 *B. sorokiniana* isolates, including *B. sorokiniana* strain DSM 62608 (GenBank Accession No. EF187908); SSU sequences had 99% homology to *Cochliobolus sativus* isolate AFTOL-ID 271 (GenBank Accession No. FJ190589).

Spot blotch caused by *B. sorokiniana* has been reported on switchgrass in Iowa, Nebraska, Pennsylvania, and Virginia (Farr and Rossman 2010). To our knowledge, this is the first report of *B. sorokiniana* causing spot blotch or common root rot of switchgrass in Tennessee, which extends the current known

distribution of these diseases. *Bipolaris sorokiniana* was isolated also from switchgrass seed received from commercial sources in the United States, indicating seedborne transmission.



Fig. 3. Conidia of *Bipolaris sorokiniana*.

***Bipolaris spicifera* (Bainier) Subramanian**

Conidiophores were single, light brown, multiseptate, mostly straight, polytretic, geniculate, and sympodial (Fig. 4A). Conidia were $17.5 \times 12 \mu\text{m}$ to $30 \times 14 \mu\text{m}$ (mean 22×12.5), oval, light brown, and distoseptate, with one to three septa and a flattened hilum on the basal cell. Conidia germinated from both poles (Fig. 4B). The causal agent was identified as *B. spicifera* (Bainier) Subram. Morphological features were as described for *B. spicifera* (Koo et al. 2003). Leaf spot symptoms similar to the original disease appeared on plants in each of the

10 inoculated replicate pots 6 days post-inoculation in the pathogenicity assay. These symptoms were not observed in the control pots. The re-isolate was also identified as *B. spicifera*. PCR amplicons of ~560 bp were obtained from both isolates and sequenced. Amplicon sequences were identical and the sequence was submitted to GenBank (Accession No. HQ015445). The DNA sequence had 100% homology to the ITS sequence of *B. spicifera* strain NRRL 47508 (GenBank Accession No. GU183125.1) that had been isolated from sorghum seed. To our knowledge, leaf spot caused by *B. spicifera* has not been described on switchgrass (Farr and Rossman 2011). *Bipolaris spicifera* can be seedborne and has been reported on turfgrass seed exported from the United States to Korea (Koo et al. 2003).

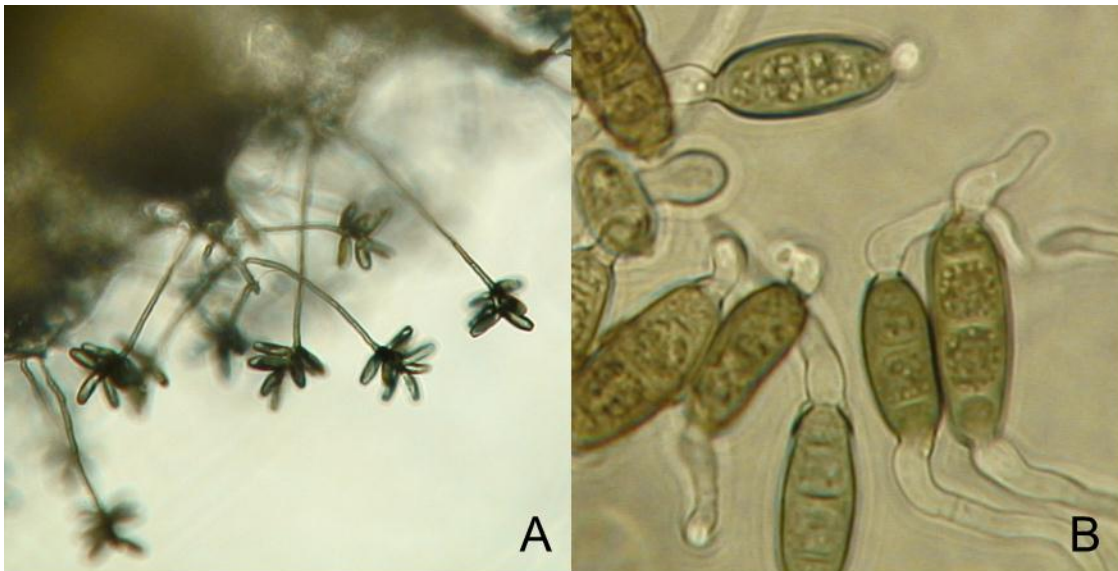


Fig 4. A. Conidiophores with conidia of *Bipolaris spicifera*, B. Bipolar germination of *B. spicifera*

***Fusarium acuminatum* Ellis & Everhart**

Cultures on PDA, incubated at 22.5 °C in the dark for 5-7 days, produced white to light yellow-orange, fluffy, aerial mycelium with sporulation. The culture was light yellow as viewed from the reverse of the culture plate. This fungus was identified as a species of *Fusarium*. After 5 days, the average colony size on PDA at 25°C in darkness was 4.5 x 4.2 cm; at 30°C with light, colony diameter was 5.2 x 4.9 cm; and at 25°C with light/20° dark conditions, the colony diameter was 5.2 x 4.8 cm. Culture color under these conditions was white to light orange-yellow. Sporodochia on CLA were light orange and spores had a foot-shaped basal cell with a slightly elongated apical cell. Spores on PDA were hyaline and boat-shaped with 3 to 5 transverse septations, with spore size ranging from 45-88 µm in length × 2.5-3.75 µm in width (Fig. 5). Chlamydospores were present after 5 to 6 weeks on CLA and no microconidia production was observed. The fungus was identified as *Fusarium acuminatum* Ellis & Everhart (Nelson et al. 1983, Booth 1971).

Inoculated plants in the pathogenicity assay developed symptoms 3 to 7 days post-inoculation for all ten replicates. Control plants had no symptoms. *Fusarium acuminatum* was re-isolated from symptomatic tissue and identified as described above.

PCR amplicons of 512 bp for the SSU region and 565 bp for the ITS region were obtained from both isolates and sequenced. Amplicon sequences were identical for the original and re-isolate for both primer sets. The SSU DNA

sequence had 99% homology to the SSU sequence of *Gibberella acuminata* (GenBank Accession No. U85550.1), the sexual stage of *Fusarium acuminatum*. The ITS DNA sequence had 99% homology to *Fusarium acuminatum* (GenBank Accession No. AB587001.1) and *Gibberella acuminata* (GenBank Accession No. U85533.1). To our knowledge, this is the first report of *F. acuminatum* on switchgrass in Tennessee, extending the known geographic range of the pathogen (Farr and Rossman 2011).



Fig. 5. Conidia of *Fusarium acuminatum*

***Fusarium equiseti* (Corda) Saccardo**

Cultures incubated in darkness at 22.5°C on PDA for 5-7 days had peach-to-light-orange, fluffy mycelium with profuse sporulation. The fungus was identified as a species of *Fusarium* and transferred to CLA to aid in species determination. Sporodochia on CLA were peach-colored and spores had a distinct foot-shaped basal cell with an elongated, tapered apical cell. Macroconidia were hyaline and boat-shaped with 4 to 7 transverse septations; spore size ranged from 27.5-88 µm in length × 2.5-5 µm in width (Fig. 6). Chlamydospores were present after several weeks on CLA. The fungus was identified as *Fusarium equiseti* (Corda) Sacc. (Nelson et al. 1983, Booth 1971).

In the pathogenicity assay, inoculated plants showed symptoms after 3 to 9 days post-inoculation for all nine replicates. Control plants showed no symptoms. The re-isolate was identified as *F. equiseti* based on morphology.

The sequences of the PCR amplicons of ~512 bp obtained from both isolates were identical and had 100% homology to several *F. equiseti* isolates in GenBank, including isolate H02-765S (GenBank Accession No. EU595566), which had been isolated from sorghum, and isolate DH08023-2 (GenBank Accession No. GU073123), isolated from green foxtail. To our knowledge, this is the first report of *F. equiseti* infecting switchgrass in Tennessee (Farr and Rossman 2011).



Fig. 6. Conidia of *Fusarium equiseti*

***Fusarium pseudograminearum* Aoki & O'Donnell**

After 5 to 7 days, the color of colonies grown on PDA and incubated at 22.5°C in the dark ranged from a deep red to pink and orange, some with white aerial mycelium and some with flat growth and sporulation (Fig.7A). Plates were vibrant pink when viewed from below. The fungus was identified as a species of *Fusarium* and transferred to CLA to aid in species determination. Spores on CLA had a foot-shaped basal cell with a tapered apical cell. Spores on PDA were hyaline, mostly straight to slightly curved with 4 to 6 transverse septations; spore size was 35-87 μm in length $\times$ 3.5-6.75 μm in width (Fig. 7B). The fungus was identified as *F. pseudograminearum* Aoki & O'Donnell (Leslie and Summerell 2006).

Symptoms were evident in inoculated plants 5 to 9 days post-inoculation for all eight replicates. Control plants showed no symptoms. *Fusarium pseudograminearum* was re-isolated from symptomatic tissue and identified on PDA as described above.

PCR amplicons of the ITS regions of approximately 525 bp were obtained from both isolates and sequenced. Amplicon sequences were identical. The DNA sequence had 99% homology to the ITS sequences of many isolates listed as “*Fusarium* sp.” DNA sequences had 99% homology also to isolates of *Fusarium graminearum*, which may be due to the fact that *F. pseudograminearum* was formerly classified as a subgroup of *F. graminearum*. Primary characteristics that differentiate these two species include disease symptomatology and whether reproduction of the isolate is hetero- or homothallic. *Fusarium graminearum* causes head blight and is heterothallic, while *F. pseudograminearum* causes crown rot and is homothallic. The isolate from switchgrass in this study was isolated from the host crown and stem. Further molecular and physiological analysis will be needed to confirm identification. To our knowledge, this is the first report of *F. pseudograminearum* causing disease on switchgrass as a pathogen, extending the known host range of the pathogen (Farr and Rossman 2011).

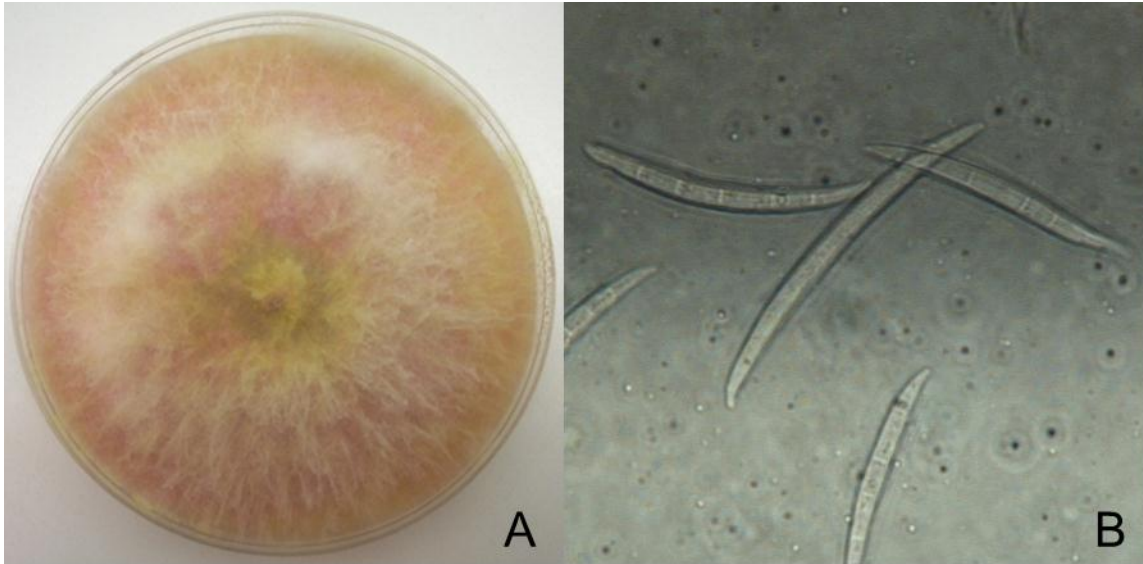


Fig. 7. A. Colony morphology of *Fusarium pseudograminearum* on potato dextrose agar, B. Conidia of *F. pseudograminearum*

***Pithomyces chartarum* (Berkeley & M.A. Curtis) M.B. Ellis**

On PDA, incubated in darkness, conidia dimensions were $10\text{--}22.5\ \mu\text{m} \times 20\text{--}37.5\ \mu\text{m}$ (average 15.17×26.5), and were golden to dark brown, broadly ellipsoidal to some pyriform, with 2 to 3 transverse septa, and a longitudinal septum constricted at the transverse septa (Fig. 8). The causal agent was identified as *Pithomyces chartarum* (Berk. & Curt.) M.B. Ellis. Morphological features were as described for *P. chartarum* (Ahonsi et al. 2010, Ellis 1971).

In the pathogenicity assay, leaf spot symptoms similar to the original disease appeared on plants in each of the eight replicate pots 6 to 10 days post-inoculation. Control plants showed no symptoms. The pathogen was re-isolated from diseased tissues and identified again as *P. chartarum*.

Sequences of the 535 bp PCR amplicons obtained from both isolates were identical. The DNA sequence had 99-100% homology to the ITS sequence of many isolates of *Leptosphaerulina chartarum* (anamorph: *Pithomyces chartarum*), including isolate Mxg-KY09-s4 (GenBank Accession No. GU195649), which had been isolated from leaf spot on *Miscanthus × giganteus* in Kentucky (Ahonsi et al. 2010). To our knowledge, leaf spot caused by *P. chartarum* has not been described on switchgrass as a host (Farr and Rossman 2011). *Pithomyces chartarum* can be seedborne and soilborne, and has been reported causing leaf spot of smooth brome grass (*Bromus inermis*) in Nebraska (Eken et al. 2006).



Fig. 8. Conidia of *Pithomyces chartarum*

***Sclerotinia homoeocarpa* F.T. Bennett**

White, fluffy, aerial mycelium developed within 12 hours of plating on PDA. Viewed from above, colonies were tan to cinnamon in color with a dark brown to black substratal stroma on and in the agar, which appeared brown viewed from below (Fig 9A). No spores were observed. Morphological characteristics of colony and hyphal growth were identical to those of *Sclerotinia homoeocarpa* F.T. Bennett (Bennett 1937).

Leaf spots appeared as early as 2 days post-inoculation in the pathogenicity assay, with full symptoms after 2 weeks for eight of nine replicates (Fig. 9B, Fig. 9C). Control plants had no symptoms.

PCR amplicons of the ITS region were ~565 bp; sequences of amplicons from the original isolate and re-isolate were identical and submitted to GenBank (Accession No. HQ850151). The ITS sequence had 99% homology with several *Sclerotinia homoeocarpa* isolates in GenBank, including three isolates from buffalograss in Oklahoma (Accession Nos. EU123800, EU123802, EU123803). The resultant 536-bp fragment was sequenced and submitted to GenBank (accession no. HQ850152); however, there were no SSU sequences from *S. homoeocarpa* available in the database for comparison. To our knowledge, this is the first confirmed report of switchgrass as a natural host for *S. homoeocarpa*, extending the known host range for the pathogen.

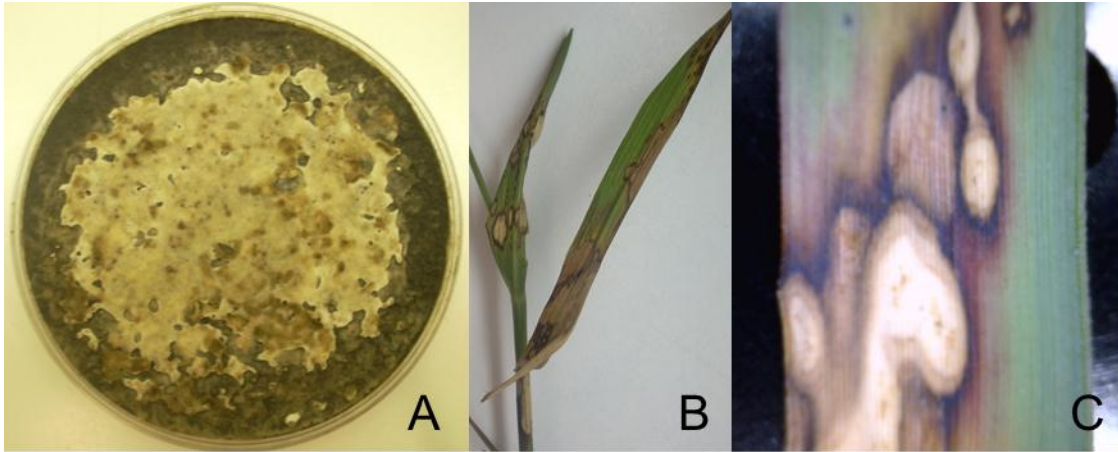


Fig. 9. A. Colony morphology of *S. homoeocarpa* on potato dextrose agar, B-C. Disease lesions caused by *Sclerotinia homoeocarpa* on switchgrass

Conclusions

Results from this study indicate that diseases caused by well-known fungal pathogens are present on switchgrass in Tennessee. These pathogens are known to cause disease in various grasses and grain crops, including various turfgrasses, miscanthus, wheat, corn, and rice. Management practices for switchgrass may need to be altered to prevent potential disease outbreaks and optimize crop yields. Several pathogens appear to have seedborne transmission. As switchgrass is increasingly grown and cultivated as a biofuels crop, efficient disease management practices and rapid disease identification procedures will likely be necessary.

References

- Ahonsi, M.O., Agindotan, B.O., Arundale, R., Gray, M.E., Voigt, T.B., and Bradley, C.A. 2010. First report of *Pithomyces chartarum* causing a leaf blight of *Miscanthus* × *giganteus* in Kentucky. Plant Dis. 94:480.
- Bennett, F.T. 1937. *Sclerotinia homoeocarpa*. Annals Appl. Biol. 24:236.
- Booth, C. 1971. The genus *Fusarium*. Commonwealth Mycological Institute: 157-179.
- Carris, L.M., Castlebury, L.A., and Zale, J. 2008. First report of *Tilletia pulcherrima* on switchgrass (*Panicum virgatum* L.) in Texas. Plant Dis. 92:1707.
- Eken, C., Jochum, C.C., Yuen, G.Y. 2006. First report of leaf spot of smooth brome grass caused by *Pithomyces chartarum* in Nebraska. Plant Dis. 90:108.
- Ellis., M. B. 1971. Dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew, Surrey, UK.
- Farr, D.F., and Rossman, A.Y. Fungal Databases, Systematic Mycology and Microbiology Laboratory ARS, USDA. Retrieved October 28, 2011.
<<http://nt.ars-grin.gov/fungaldatabases/>>
- Gravert, C.E. and Munkvold, G.P. 2002. Fungi and diseases associated with cultivated switchgrass in Iowa. J. Iowa Acad. Sci. 109:30-34.

- Gwinn, K.D., Trigliano, R.N., Gavin, A.M., and Conger, B.V. 1991. Bacterial interference with in vitro assays of tall fescue seeds for *Acremonium coenophialum*. Crop Sci. 31:1369-1370.
- Koo, H.M., Lee, S.H., Chung, I.M., and Chun, S.E. 2004. Cultural characteristics of a seedborne fungus, *Bipolaris spicifera* detected from imported grass seeds into Korea. Mycobiology 32:186-189.
- Krupinsky, J.M., Berdahl, J.D., Schoch, C.L., and Rossman, A.Y. 2004. Leaf spot on switchgrass (*Panicum virgatum*), symptoms of a new disease caused by *Bipolaris oryzae*. Can. J. Pl. Pathol. 26:371-378.
- Leslie, J.F., and Summerell, B.A. 2006. The Fusarium Laboratory Manual. Blackwell Publishing. Ames, Iowa.
- Li, K.N., Rouse, D.I., and German, T.L. 1994. PCR primers that allow intergeneric differentiation of ascomycetes and their application to *Verticillium* spp. Appl. Environ. Microbiol. 60:4324-4331.
- Nelson, P.E., Toussoun, T.A., and Marassas. 1983. *Fusarium* species, an illustrated manual for identification. Pennsylvania State University, University Park and London: 89-100.
- Nyvall, R. F. and Percich, J. A. 1999. Development of fungal brown spot and spot blotch on cultivated wild rice in Minnesota. Plant Dis. 83:936-938.
- Schmer, M.R., Vogel, K.P., Mitchell, R.B., and Perrin, R.K. 2008. Net energy of cellulosic ethanol from switchgrass. PNAS 105:464-469.

- Shahin, E.A., and Shephard, J.F. 1979. An efficient technique for inducing profuse sporulation of *Alternaria* species. *Phytopathology* 69:618-620.
- Sivanesan, A. and P. Holliday. 1981. *Cochliobolus sativus*. CMI Descr. Pathog. Fungi bact. 71:701.
- Sivanesan, A. 1987. Graminicolous species of *Bipolaris*, *Curvularia*, *Drechslera*, *Exserohilum* and their teleomorphs, *Mycolog. Pap.* 158:201.
- Tomaso-Peterson, M., and Balbalian, C.J. 2010. First report of *Bipolaris oryzae* causing leaf spot of switchgrass in Mississippi. *Plant Dis.* 94:643.
- White, T.J. 1990. Page 315 in: *PCR Protocols: A Guide to Methods and Applications*. M. A. Innis et al., eds, Acad. Press, NY.
- Zale, J., Freshour, L., Agarwal, S., Sorochan, J., Ownley, B.H., Gwinn, K.D., Castlebury, L.A., and Carris, L.M. 2008. First report of *Puccinia emaculata* on switchgrass (*Panicum virgatum* L.) in Tennessee. *Plant Dis.* 92:1710.

CHAPTER 3

ASSESSMENT OF PATHOGENS IN SWITCHGRASS SEED PRODUCED IN THE UNITED STATES

Abstract

Switchgrass (*Panicum virgatum* L.) production is increasing in acreage due to current initiatives for commercial biofuel production in the United States and worldwide. Due to a lack of seed certification programs for switchgrass, seedborne plant pathogens have likely been shipped along with the seeds to switchgrass producers. Management of seedborne pathogens likely would increase stand establishment and crop yields, while decreasing the likelihood of a seedborne epidemic.

The aim of this study was to determine if seedborne fungal pathogens were present in switchgrass seed commercially produced in the U.S., and if present, their identity and frequency. Seed of seven cultivars from 11 sources were tested, including multiple entries of 'Alamo', 'Blackwell', 'Cave-in-Rock', and 'Kanlow'. A randomly-selected subsample of seed from each 454-g seed lot was surface-sterilized in 1% NaOCl for 1 min, rinsed three times with sterile water, and dried on sterile filter paper. Three hundred surface-sterilized seed per lot were plated on potato dextrose agar (PDA) amended with 100 mg/liter chloramphenicol and incubated at 22°C. Seed were evaluated daily for development of fungal colonies. Rates of fungal infection among the 30 sampled seed lots ranged from less than 1 % to 87%. The most frequently evaluated pathogens were *Bipolaris oryzae*, *Alternaria alternata*, and *Fusarium graminearum/pseudograminearum*. Additional species of *Bipolaris* and *Fusarium* were present, but in lower frequency.

Introduction

Switchgrass (*Panicum virgatum* L.) is a warm-season grass that, due to its ability to adapt to a wide variety of environmental conditions, low fertility requirements, and low production cost, is being grown for biofuel production across the US and around the world. Although switchgrass is a perennial plant, stand establishment has proven to be a problem for growers. One factor contributing to the problem of stand establishment is poor seed quality (Parrish and Fike 2005; Sanderson et al. 2006). To our knowledge, only two studies have been conducted to examine switchgrass seed for pathogens (Carris et al. 2008, Tomaso-Peterson and Balbalian 2010).

In our studies, we have observed increased stand establishment and enhanced growth and vigor with surface-sterilized seed versus untreated seed (unpublished data). Currently, no seed certification program exists for switchgrass. The objective of this study was to identify major seedborne pathogens, their incidence in diverse seed lots, and the current geographic distribution of these organisms as determined by seed source location.

Materials and Methods

Thirty switchgrass seed lots, including cultivars 'Alamo', 'Blackwell', 'Cave-in-Rock', 'Forestburg', 'Kanlow', 'Cultivar X', and 'Cultivar Y', were obtained from eleven companies in the U.S (Table 1). Since 'Cultivar X' and 'Cultivar Y' are only

available from one company each, the names have been change to prevent interference with the ability of the companies to sell their products. Company names were changed for the same reason.

Table 1. Cultivars and source of switchgrass seed

Cultivar	Company	Company Location	Seed Source
'Alamo'	Company 1	CO	TX
	Company 2	NM	TX
	Company 3	PA	MO
	Company 5	OK	OK
	Company 6	OK	OK
	Company 7	KY	TN
	Company 8	KS/ CO	OK
	Company 9	TX	TX
	Company 10	OK	OK
'Blackwell'	Company 1	CO	OK
	Company 2	NM	SD
	Company 3	PA	PA
	Company 5	OK	SD
	Company 7	KY	TN
	Company 8	KS/ CO	KS
	Company 9	TX	TX
	Company 10	OK	TX
'Cave-in-Rock'	Company 1	CO	IL
	Company 3	PA	PA
	Company 7	KY	KY
	Company 8	KS/ CO	KS
	Company 11	IA	IA
'Kanlow'	Company 1	CO	OK
	Company 3	PA	IA
	Company 5	OK	OK
	Company 7	KY	MO
	Company 8	KS/ CO	MO
'Forestburg'	Company 4	WI	CO
'Cultivar X'	Company 5	OK	OK
'Cultivar Y'	Company 6	OK	OK

A randomly selected subset of seeds from each lot was surface-disinfested with 1% NaOCl (Clorox, Oakland, CA) amended with 10 µl Tween 20 (U. S. Biochemical Corp., Cleveland, OH) for 1 min with agitation, rinsed three times with sterile deionized water, and dried on sterile filter paper (Mathur and Kongsdal 2003). Three hundred seeds per lot, at a rate of 25 seeds per 9-cm diameter Petri dish, were plated on potato dextrose agar (PDA) amended with 100 mg chloramphenicol/liter of PDA (Sigma-Aldrich, St. Louis, MO). Seeds were checked daily for fungal growth. Emergent colonies were transferred to individual PDA plates for identification. Colonies were identified by colony morphology, spore characteristics, and internal transcribed spacer (ITS) region of 18S ribosomal DNA sequences.

Simpson's index of diversity was calculated for each seed lot (Peet 1974). Simpson's index of diversity is the percent likelihood that two organisms randomly picked from the total population are of different species, thus taking into account species richness and abundance.

Results and Discussion

Nine thousand switchgrass seed were examined for fungal colonization and 2,280 isolates were recovered from these surface-sterilized seed. Of the fungal species recovered, many have been reported as pathogens of switchgrass seedlings or mature plants, including *Alternaria alternata*, *Bipolaris oryzae*, *B.*

sorokiniana, *B. spicifera*, *Curvularia* spp., *Fusarium equiseti*, *F. graminearum*/*pseudograminearum* and *F. oxysporum* (Farr and Rossman 2011). In this study, *A. alternata*, *B. oryzae*, and *F. graminearum*/*pseudograminearum* were the most commonly isolated pathogenic fungi, with infection rates as high as 38.3, 34.7, and 20.7%, respectively, in some seed lots (Table 2). Several other pathogenic *Bipolaris* species, including *B. sorokiniana*, *B. spicifera*, and *B. victoriae*, and *Fusarium* species, such as *F. equiseti*, were found with less frequency. Overall seedlot infection rates ranged from 0.7-86.8% (Table 3), taking into account that both pathogenic and non-pathogenic fungi were present in some instances.

Table 2. Percent infection of switchgrass seed by cultivar, source and fungal species

Cultivar	Source	Fungal species					
		<i>Alternaria alternata</i>	<i>Aspergillus</i> spp.	<i>Bipolaris oryzae</i>	<i>Bipolaris sorokiniana</i>	<i>Bipolaris spicifera</i>	
'Alamo'	Company 1	3.0	0.0	0.3	0.0	0.0	
	Company 2	4.0	0.0	1.0	0.0	0.0	
	Company 3	6.0	0.0	1.0	0.0	0.0	
	Company 5	0.7	0.0	0.5	0.7	0.0	
	Company 6	13.0	0.0	0.0	0.0	0.3	
	Company 7	2.7	0.0	0.3	0.0	0.0	
	Company 8	1.3	13.0	0.0	0.0	0.0	
	Company 9	0.0	0.0	0.0	0.0	0.0	
	Company 10	0.0	0.0	0.0	0.0	0.0	
	'Blackwell'	Company 1	5.7	0.0	0.0	0.3	0.0
Company 2		1.7	0.0	0.3	0.0	0.0	
Company 3		0.0	0.0	0.0	0.0	0.0	
Company 5		0.0	0.0	0.0	0.0	0.0	
Company 7		3.7	0.0	0.7	0.0	0.0	
Company 8		29.3	0.0	0.0	0.3	0.0	
Company 9		38.3	0.0	0.0	0.0	0.0	
Company 10		0.0	0.0	14.7	0.0	0.0	
'Cave-in-Rock'		Company 1	20.3	0.0	29.7	1.7	0.0
		Company 3	9.5	0.0	3.5	0.0	0.0
	Company 7	8.7	0.0	0.7	0.0	0.0	
	Company 8	38.3	0.0	0.0	0.0	0.0	
	Company 11	11.3	0.0	34.7	0.3	0.0	
'Kanlow'	Company 1	13.3	0.0	7.7	0.0	0.0	
	Company 3	0.0	0.0	0.0	0.0	0.0	
	Company 5	0.0	0.0	0.0	0.0	0.0	
	Company 7	5.7	0.0	2.0	0.0	0.0	
	Company 8	22.5	0.0	20.0	0.3	0.0	
'Forestburg'	Company 4	0.0	0.0	0.0	0.0	0.0	
'Cultivar X'	Company 5	31.8	0.0	0.0	0.0	0.0	
'Cultivar Y'	Company 6	0.7	0.0	0.0	0.0	0.0	

Table 2. Percent infection of switchgrass seed by cultivar, source and fungal species (continued)

		Fungal species					
Cultivar	Source	<i>Bipolaris victoriae</i>	<i>Bipolaris zeicola</i>	<i>Chaetomium globosum</i>	<i>Curvularia</i> spp.	<i>Eurotium</i> sp.	
'Alamo'	Company 1	0.0	0.0	0.0	0.0	0.7	
	Company 2	0.0	0.0	0.0	0.0	0.0	
	Company 3	0.0	0.0	0.0	0.0	0.0	
	Company 5	0.0	0.3	10.3	0.0	1.5	
	Company 6	0.0	0.0	0.0	0.0	0.0	
	Company 7	0.0	0.0	0.0	0.0	0.0	
	Company 8	0.0	0.0	0.0	0.0	0.0	
	Company 9	0.0	0.0	3.7	0.0	1.3	
	Company 10	0.0	0.0	0.7	0.0	0.0	
	'Blackwell'	Company 1	0.0	0.0	0.0	0.0	0.0
Company 2		0.0	0.0	1.3	0.0	0.0	
Company 3		0.0	0.0	0.0	0.0	1.3	
Company 5		0.0	0.0	0.3	0.0	0.0	
Company 7		0.0	0.0	0.0	0.0	0.0	
Company 8		0.0	0.0	0.0	0.0	0.0	
Company 9		0.0	0.0	0.0	0.0	0.0	
Company 10		0.0	0.0	0.0	0.0	0.0	
'Cave-in-Rock'		Company 1	0.2	0.0	0.0	0.0	0.0
		Company 3	0.0	0.0	0.0	0.0	0.0
	Company 7	0.0	0.0	0.3	0.0	0.0	
	Company 8	0.0	0.0	0.0	0.0	0.0	
	Company 11	0.3	0.0	0.0	0.0	0.0	
'Kanlow'	Company 1	0.3	0.0	0.0	0.3	0.0	
	Company 3	0.0	0.0	0.0	0.0	0.0	
	Company 5	0.0	0.0	3.8	0.0	8.8	
	Company 7	0.0	0.0	3.7	0.0	3.3	
	Company 8	0.3	0.0	0.0	0.3	0.0	
'Forestburg'	Company 4	0.0	0.0	4.0	0.3	0.0	
'Cultivar X'	Company 5	0.0	0.0	1.0	0.3	0.0	
'Cultivar Y'	Company 6	0.0	0.0	0.0	0.0	0.0	

Table 2. Percent infection of switchgrass seed by cultivar, source and fungal species (continued)

Cultivar	Source	Fungal species				
		<i>Fusarium equiseti</i>	<i>Fusarium graminearum/ pseudo-graminearum</i>	<i>Fusarium oxysporum</i>	<i>Fusarium</i> spp.	<i>Penicillium</i> spp.
'Alamo'						
	Company 1	0.0	0.0	0.0	0.0	0.0
	Company 2	0.0	0.3	0.0	0.7	0.0
	Company 3	2.7	7.7	0.0	7.3	0.3
	Company 5	0.0	0.0	0.0	0.0	0.0
	Company 6	0.0	0.0	0.0	0.0	0.0
	Company 7	0.0	0.2	4.5	31.3	0.7
	Company 8	0.0	0.0	0.0	0.0	3.0
	Company 9	0.0	0.7	0.0	0.0	0.0
	Company 10	0.0	0.0	0.0	0.0	0.0
'Blackwell'						
	Company 1	2.7	0.0	0.0	0.7	0.0
	Company 2	0.0	0.7	0.0	1.3	0.3
	Company 3	0.0	0.0	0.0	1.0	2.7
	Company 5	0.0	0.0	0.0	0.0	0.7
	Company 7	0.0	1.2	0.0	4.0	0.3
	Company 8	0.0	0.0	0.0	0.0	0.0
	Company 9	0.0	1.5	0.0	0.0	0.0
	Company 10	0.0	0.0	0.0	0.0	0.0
'Cave-in-Rock'						
	Company 1	0.0	0.0	5.0	1.0	0.0
	Company 3	0.0	20.7	0.0	12.8	0.0
	Company 7	0.0	0.0	0.0	1.0	0.7
	Company 8	0.0	0.0	0.0	0.7	0.0
	Company 11	0.0	0.0	0.0	0.0	0.3
'Kanlow'						
	Company 1	0.0	0.0	0.0	10.7	0.0
	Company 3	0.0	0.0	0.0	0.0	0.0
	Company 5	0.0	0.0	0.0	0.0	0.0
	Company 7	0.0	0.3	0.0	7.0	0.0
	Company 8	0.0	4.0	0.0	12.7	2.5
'Forestburg'						
	Company 4	0.0	0.0	0.0	1.7	0.0
'Cultivar X'						
	Company 5	0.0	4.7	0.0	8.5	0.0
'Cultivar Y'						
	Company 6	0.0	0.0	0.0	0.0	0.0

Table 2. Percent infection of switchgrass seed by cultivar, source and fungal species (continued)

Cultivar	Source	Fungal species					
		<i>Phoma</i> sp.	<i>Pithomyces</i> <i>chartarum</i>	<i>Xylaria</i> sp.	Zygo- mycetes	Unidentified brown Ascomycete	Un- Identified fungi
'Alamo'							
	Company 1	0.0	0.0	0.3	0.0	0.0	0.0
	Company 2	1.7	0.0	0.0	0.0	0.0	1.0
	Company 3	2.0	0.0	0.0	0.0	0.0	2.3
	Company 5	0.0	0.0	0.0	0.0	0.0	0.0
	Company 6	2.0	0.0	0.7	0.0	0.0	0.7
	Company 7	0.0	0.0	0.0	0.0	0.0	0.0
	Company 8	0.7	0.0	0.3	7.7	0.0	0.7
	Company 9	0.0	0.0	0.0	0.0	0.0	0.0
	Company 10	0.3	0.0	0.0	0.0	0.0	1.7
'Blackwell'							
	Company 1	0.0	0.0	0.0	0.0	0.0	2.3
	Company 2	0.3	0.0	0.7		6.3	0.0
	Company 3	2.0	0.0	0.0	0.0	0.0	2.0
	Company 5	0.0	0.0	0.0	0.0	0.0	0.3
	Company 7	12.8	0.0	0.0	0.0	0.0	1.0
	Company 8	0.0	0.0	0.3	0.0	0.0	1.0
	Company 9	0.3	0.0	4.2	0.0	0.0	0.0
	Company 10	1.0	0.0	0.0	0.0	0.0	0.3
'Cave-in-Rock'							
	Company 1	25.5	0.0	1.2	0.0	0.0	2.3
	Company 3	0.0	0.3	0.2	0.0	0.0	5.7
	Company 7	0.3	0.0	0.0	2.0	0.0	1.7
	Company 8	0.3	0.0	0.3	0.0	0.0	0.0
	Company 11	4.7	0.0	0.0	0.0	0.0	0.0
'Kanlow'							
	Company 1	0.0	0.0	0.0	0.0	0.0	0.0
	Company 3	0.0	0.0	0.0	0.0	0.0	0.7
	Company 5	0.0	0.0	0.0	0.0	0.0	0.0
	Company 7	3.7	0.0	0.0	0.0	0.0	0.0
	Company 8	7.0	2.5	0.3	0.0	0.0	6.7
'Forestburg'							
	Company 4	0.0	0.0	0.0	6.0	0.3	0.0
'Cultivar X'							
	Company 5	3.7	0.0	0.3	0.0	0.0	0.3
'Cultivar Y'							
	Company 6	0.0	0.0	0.0	0.0	0.0	0.7

Table 3. Total percent switchgrass seed infection and diversity analysis by cultivar and source

Cultivar	Source	Total percent infection	Total percent <i>Bipolaris</i> spp. infection	Total percent <i>Fusarium</i> spp. infection	Simpson's Index of Diversity ¹
'Alamo'					
	Company 1	4.3	0.3	0.0	52.56
	Company 2	8.7	1.0	1.0	77.78
	Company 3	29.3	1.0	17.7	81.61
	Company 5	14.0	1.5	0.0	44.8
	Company 6	16.7	0.3	0.0	38.12
	Company 7	39.7	0.3	36.0	36.13
	Company 8	26.7	0.0	0.0	67.15
	Company 9	5.7	0.0	0.7	54.41
	Company 10	2.7	0.0	0.0	60.71
'Blackwell'					
	Company 1	11.7	0.3	3.3	68.74
	Company 2	13.0	0.3	2.0	73.68
	Company 3	9.0	0.0	1.0	80.91
	Company 5	1.3	0.0	0.0	66.67
	Company 7	23.7	0.7	5.2	65.74
	Company 8	31.0	0.3	0.0	8.48
	Company 9	44.3	0.0	1.5	24.42
	Company 10	16.0	14.7	0.0	15.87
'Cave-in-Rock'					
	Company 1	86.8	31.5	6.0	74.03
	Company 3	52.7	3.5	33.5	74.28
	Company 7	15.3	0.7	1.0	64.14
	Company 8	39.7	0.0	0.7	6.62
	Company 11	51.7	35.3	0.0	49.66
'Kanlow'					
	Company 1	32.3	8.0	10.7	67.16
	Company 3	0.7	0.0	0.0	0
	Company 5	12.7	0.0	0.0	43.35
	Company 7	25.7	2.0	7.3	82.37
	Company 8	79.2	20.7	16.7	81.37
'Forestburg'					
	Company 4	12.3	0.0	1.7	65.62
'Cultivar X'					
	Company 5	50.7	0.0	13.2	56.66
'Cultivar Y'					
	Company 6	1.3	0.0	0.0	66.67

¹ Simpson's Index of Diversity= $1-(\sum_{i=1}^S P_i^2)$, where P_i = proportion of all individuals belonging to the i^{th} species

In this study, several fungal pathogens of switchgrass that were not previously shown to have seedborne transmission were isolated directly from seed. 'Cave-in-Rock' appears to be more susceptible to seedborne infection with the known pathogens, but statistical analysis was not performed. Although no direct correlation between cultivar, company, or seed source location and percent infection by the known pathogens was determined, the information provided by this study may help to prevent the seedborne movement of pathogens around the country and around the world, and explain the movement of switchgrass pathogens, such as *B. oryzae*. *Bipolaris oryzae*, which was found in 50% of the seed lots tested, including cultivars 'Alamo', 'Blackwell', 'Cave-in-Rock', and 'Kanlow' (Table 2), has been reported from many locations around the United States, including New York, North Dakota, Mississippi, and Oklahoma. Some of these reports have been descriptions of severe outbreaks (Waxman and Bergstrom 2011, Krupinsky et al. 2004, Tomaso-Peterson et al. 2010, and Ghimire et al. 2011). Further studies will be needed to determine/confirm species identity and pathogenicity of several fungi isolated from seed, including *Phoma* sp., *Curvularia* spp., *Fusarium* spp., *Bipolaris victoriae*, and *Bipolaris zeicola*.

The most frequently isolated species were *Alternaria alternata*. This pathogen has a broad host range, including over 900 host species (Farr and Rossman 2011). *Alternaria alternata* can cause disease on wheat and rice, in addition to inducing problems in humans as an environmental allergen causing chronic sinusitis and as a cause of esophageal cancer when the mycotoxins

produced on grains (including wheat) are ingested (Liu et al. 1991, Liu et al. 1992). This fungus can survive for a long time as a saprophyte in soil, and mycotoxin production may prove problematic for bioethanol conversion from switchgrass infected with *A. alternata*.

Another large group of fungi identified are the anamorphic stages of *Cochliobolus*: *Bipolaris* and *Curvularia*. Both genera contain pathogens of a range of grass hosts, including turfgrasses, sorghum, wheat, corn, and oat. These pathogens frequently can survive in soil, which may need to be taken into account for crop rotations, particularly in the southeastern U. S.

Several species of *Fusarium* were isolated from seed in this study, and more work needs to be done to confirm species identity. Several of the species also were identified from mature plants grown in fields or in ornamental plots in Tennessee in previous work, including *Fusarium graminearum*/*pseudograminearum* and *Fusarium equiseti* (See Chapter 2 of this thesis). Both *F. graminearum*/*pseudograminearum* and *F. equiseti* are likely to cause disease problems in switchgrass necessitating the development of management practices. Species of *Fusarium* can also produce mycotoxins that could interfere with the biomass conversion process, which uses microbial and enzymatic digestion and fermentation of sugars to produce ethanol. Further studies will need to be conducted to assess the effect of mycotoxin presence on overall bioethanol yield.

Another well-known mycotoxin producer found in this study was *Pithomyces chartarum*, which can also causes diseases of sheep and other livestock. *Pithomyces chartarum* was not found frequently in this study, but had previously been isolated from switchgrass grown in a growth chamber for pot studies (See Chapter 2 of this thesis). *Phoma* sp. also was reported previously on switchgrass, but pathogenicity was not determined (Farr and Rossman 2011). *Phoma* can cause diseases on related grasses, such as wild rice (Nyvall and Percich 1999).

Species of *Xylaria*, *Eurotium*, and *Chaetomium* are likely to be non-pathogenic endophytes. More studies will be needed to determine if any of these fungi have an effect on growth of switchgrass. When measures are taken to decrease overall fungal infection of seed, presence or absence of species of these genera may not have any effect on overall switchgrass growth or stand establishment.

More work will need to be conducted to determine the best management plan for these pathogens. Likely, decreasing inoculum levels in the field will have a profound effect on overall seed infection rates. Seed treatments may be explored as an effective control method if germination and emergence problems become limiting factors in establishing a quality stand.

References

- Carris, L.M., Castlebury, L.A., and Zale, J. 2008. First report of *Tilletia pulcherrima* bunt on switchgrass (*Panicum virgatum*) in Texas. Plant Dis. 92:1707.
- Farr, D.F., and Rossman, A.Y. 2011. Fungal Databases, Systematic Mycology and Microbiology Laboratory, ARS, USDA. Retrieved August 4, 2011, from <http://nt.ars-grin.gov/fungaldatabases/>.
- Ghimire, S.R., Charlton, N.D., Bell, J.D., Krishnamurthy, Y.L., and Craven, K.D. 2011. Biodiversity of fungal endophyte communities inhabiting switchgrass (*Panicum virgatum* L.) growing in the native tallgrass prairie of northern Oklahoma. Fung. Diversity 47:19-27.
- Krupinsky, J.M., Berdahl, J.D., Schoch, C.L., and Rossman, A.Y. 2004. Leaf spot on switchgrass (*Panicum virgatum*), symptoms of a new disease caused by *Bipolaris oryzae*. Can. J. Plant Pathol. 26:371-378.
- Liu, G.T., Qian, Y.Z., Zhang, P., Dong, W.H., Qi, Y.M., and Guo, H.T. 1992. Etiological role of *Alternaria alternata* in human esophageal cancer. Chin. Med J. 105:394-400.
- Liu, G.T., Qian, Y.Z., Zhang, P., Dong, Z.M., Shi, Z.Y., Zhen, Y.Z., Miao, J., and Xu, Y.M. 1991. Relationships between *Alternaria alternata* and oesophageal cancer. IARC Sci Publ. 105:258-262.

- Mathur, S.B., and Kongsdal, O. 2003. Common Laboratory Seed Health Testing Methods for Detecting Fungi. International Seed Testing Association, Basserdorf, CH-Switzerland.
- Nyvall, R. F. and Percich, J. A. 1999. Development of fungal brown spot and spot blotch on cultivated wild rice in Minnesota. *Plant Dis.* 83:936-938.
- Parrish, D.J. and Fike, J.H. 2005. The biology and agronomy of switchgrass for biofuels. *Crit. Rev. Plant Sci.* 24:423–459.
- Peet, R.K. 1974. The measurement of species diversity. *Ann. Rev. Ecol. System.* 5:285-307.
- Sanderson, M.A., Adler, P.R., Boateng, A.A., Casler, M.D. and Sarath, G. 2006. Switchgrass as a biofuels feedstock in the USA. *Can. J. Plant Sci.* 86:1315–1325.
- Tomaso-Peterson, M., and Balbalian, C.J. 2010. First report of *Bipolaris oryzae* causing leaf spot of switchgrass in Mississippi. *Plant Dis.* 94:643.
- Waxman, K.D., and Bergstrom, G.C. 2011. First report of a leaf spot caused by *Bipolaris oryzae* on switchgrass. *Plant Dis.* 95:1192.

CHAPTER 4

**INVESTIGATING ANTIMICROBIAL ACTIVITY OF
SWITCHGRASS-DERIVED EXTRACTIVES**

Abstract

Switchgrass-derived ethanol is currently being investigated as a renewable energy source alternative to fossil fuel. However, conversion of switchgrass biomass to ethanol is still being optimized. Previous studies have shown that switchgrass-derived extractives can contain phenolic compounds associated with antimicrobial activity. Due to the microbial and enzymatic involvement in many systems for saccharification and fermentation of plant material in the conversion to ethanol, antimicrobial activity could decrease overall ethanol yield. In this study, switchgrass extractives were evaluated for antimicrobial activity. Six ethanol-soluble extractives treatments, extracted from switchgrass collected from three farms at two different plant ages, were tested in a 3×2 factorial design against four bacterial plant pathogens: *Clavibacter michiganensis* subsp. *michiganensis*, *Xanthomonas perforans*, *Pseudomonas syringae* pv. *tomato*, and *Pseudomonas mediterranea*. Significant differences were shown for the main effect of bacteria ($P=0.09$) and the interaction effect of farm $\times$ plant age ($P=0.04$). *Xanthomonas perforans* was more sensitive to the extractives treatments than *Pseudomonas syringae* pv. *tomato* and extractives from 112 day-old switchgrass from farm C04 inhibited bacterial growth more than the extractives from 112 day-old switchgrass from farms C19 and C33. There was no difference in bacterial inhibition among farms for the extractives from 56 day-old switchgrass plants. In addition to increasing the overall ethanol yield per gram of plant tissue, removal of the extractives and use of these compounds as

an antimicrobial agent will lead to the use of the extractives as a value-added product to the biomass conversion system.

Introduction

Switchgrass (*Panicum virgatum* L.) is a perennial warm-season (C₄) prairie grass that is native to many regions of the United States. Switchgrass is being investigated as a lignocellulose feedstock crop due to its wide adaptability and perennial growth characteristics, but the process of breaking down switchgrass plant material to fermentable sugars is still being optimized. Several systems have been developed that use mechanical/chemical disruption to breakdown complex sugars to simpler sugars through a pretreatment step prior to saccharification and fermentation (Tao et al. 2011). Saccharification and fermentation employ a battery of microorganisms for sugar breakdown and conversion (Keshwani and Cheng 2009). When switchgrass plant material is ground and extracted with polar solvents, extractives (low molecular-weight lipophilic and hydrophilic compounds) are released into solution, leaving behind desired carbohydrates. Ethanol-soluble extractives contain various fatty acids, fatty alcohols, glycerol, alkanes, and sterols (Yan et al. 2010). When plant material is extracted with hot water, composition of extractives is similar to that of the ethanol-soluble extractives (Thammasouk et al. 1997). Hot water extractives contain antioxidants, such as α -tocopherol and policosanol (Ravindranath et al. 2009). Flavonoids, such as quercitrin and rutin, can be found in the compounds

extracted with both hot water and methanol (Uppugundla et al. 2009). Water-soluble extractives from switchgrass were shown to be composed of 30 to 35% of a “red-brown” fraction, which in experiments with corn stover conducted by the same research laboratory contained phenolic compounds such as *p*-coumaric, ferulic, and sinapic acids (Chen et al. 2007, Chen et al. 2010). The solubility of these compounds into solution during saccharification and fermentation could potentially inhibit enzymatic and microbial activity, thereby, decreasing overall ethanol yields per gram of starting material. Removal of the extractives prior to the pretreatment step may increase overall conversion efficiency, and the extractives could be exploited for antimicrobial activity as a value-added product. In this study, total phenols were measured and the antimicrobial activity of switchgrass-derived extractives was tested against four bacterial plant pathogens: *Clavibacter michiganensis* subsp. *michiganensis*, *Xanthomonas perforans*, *Pseudomonas syringae* pv. *tomato*, and *Pseudomonas mediterranea*.

Materials and Methods

Bacteria

Four plant pathogenic bacteria were tested for susceptibility to antimicrobial activity by switchgrass-derived extractives: *Clavibacter michiganensis* subsp. *michiganensis*, *Xanthomonas perforans*, *Pseudomonas syringae* pv. *tomato*, and *Pseudomonas mediterranea*. All species were from culture collections maintained by Dr. Bonnie Ownley (University of Tennessee) and are known to be pathogenic on tomato. *Clavibacter michiganensis* subsp. *michiganensis* (CMM), *Xanthomonas perforans* (XS), and *Pseudomonas syringae* pv. *tomato* (PST) were isolated from tomato and *Pseudomonas mediterranea* (PM) was isolated from soybean. Each culture was shaken overnight in tryptic soy broth. Cultures were centrifuged for 3 min at $4,200 \times g$ in an Eppendorf 5810 centrifuge (Eppendorf AG, Hamburg, Germany). Broth was then removed and each culture was resuspended in phosphate buffered saline (PBS) at a concentration of approximately 1×10^8 CFU/ml.

Extractives

Six groups of 'Alamo' switchgrass were harvested from three different farms in Vonore, TN at two different plant ages (Table 1). The extractives from each sample were separated with 95% ethanol (Decon Laboratory, Inc, PA, USA) using a Dionex Accelerated Solvent Extractor (Dionex, Sunnyvale, CA).

Ethanol was driven off with an HBA 10 Digital Rotary Evaporator (IKA, Germany) and Model 281A Vacuum Oven (Fisher Scientific, PA, USA). This work was conducted by Dr. Paul Filson and Britany Swann. Samples were stored at 4°C in darkness until experiments were performed.

For each extractives sample, 125 mg of dry extractives material was dissolved in 4 ml of 10% ethanol solution, vortexed, sonicated for three 1-min intervals, and cold-sterilized with a 0.2-µm filter into a sterile glass vial. Total phenolics were measured following the Folin-Ciocalteu analysis procedure (Waterhouse 2002) and with a UV160U UV-visible recording spectrophotometer (Shimadzu Corporation, Kyoto, Japan).

Table 1. Source and plant age of six ‘Alamo’ selections from which extractives were obtained

Farm	Plant age (in days)
C04	56
C04	112
C19	56
C19	112
C33	56
C33	112

Treatments

Antimicrobial activity of extractives was tested by making a 50% (v/v) solution of 0.5 ml extractives solution and 0.5 ml bacterial solution in PBS for each of the six extractives, and one control, for each bacterial isolate. These solutions were incubated at 22°C on the laboratory benchtop for 60 min. Solutions were then diluted with PBS to 10^{-5} , 10^{-6} , and 10^{-7} of the original dilution and plated on tryptic soy agar in duplicate. Plates were counted and dilution plates with colony counts in the range of 30 to 300 were used for data analysis. The experiment was replicated over time with three trials, and one replicate per trial. The mean of population counts of two dilution plates was used for each replicate.

Experimental Design and Statistical Analysis

The experimental setup was a 3×2 factorial with three farm locations and two plant ages. The variable $\text{Log}_{10}(\text{Treatment Population Count} + 1)$ was calculated for each combination of bacteria and extractives, and bacteria and control. Population growth was determined as a percentage of control:

$$\frac{\text{Log}_{10}(\text{Treatment Population Count} + 1)}{\text{Log}_{10}(\text{Control Population Count} + 1)} \times 100$$

The main effects and interactions of extractives and bacteria on this variable were analyzed for significance with the Proc Mixed procedure of SAS 9.2 software (SAS Institute, Cary, NC). Significant effects and interactions were

further analyzed with a Fisher's-protected least significant difference (F-LSD) test at $P=0.05$ and $P=0.10$.

Results and Discussion

Total phenols

Total phenolic concentrations varied from 7.19 to 21.06 mg GAE per g dry extractives (Table 2).

Table 2. Total phenolic concentrations

Farm	Plant age (in days)	mg GAE ¹ /g extractives
C04	56	7.19 ± 0.97
C04	112	12.17 ± 1.00
C19	56	13.81 ± 5.49
C19	112	12.45 ± 0.93
C33	56	21.06 ± 2.92
C33	112	8.03 ± 0.87

¹GAE = Gallic acid equivalent. Values are the mean of three replicates ± standard error.

Bacterial inhibition

The main effect of bacteria ($P = 0.0967$) and the interaction of farm $\times$ plant age ($P = 0.0494$) were significant for population growth as a percentage of control. *Xanthomonas perforans* was significantly more sensitive to the extractives treatments overall than *Pseudomonas syringae* pv. *tomato* (Fig. 1). A significant difference was also shown between farm C04 and farm C19 and C33 for plant age = 112 days, in which switchgrass extractives from farm C04 had more antimicrobial activity than extractives from C19 and C33 (Fig. 2).

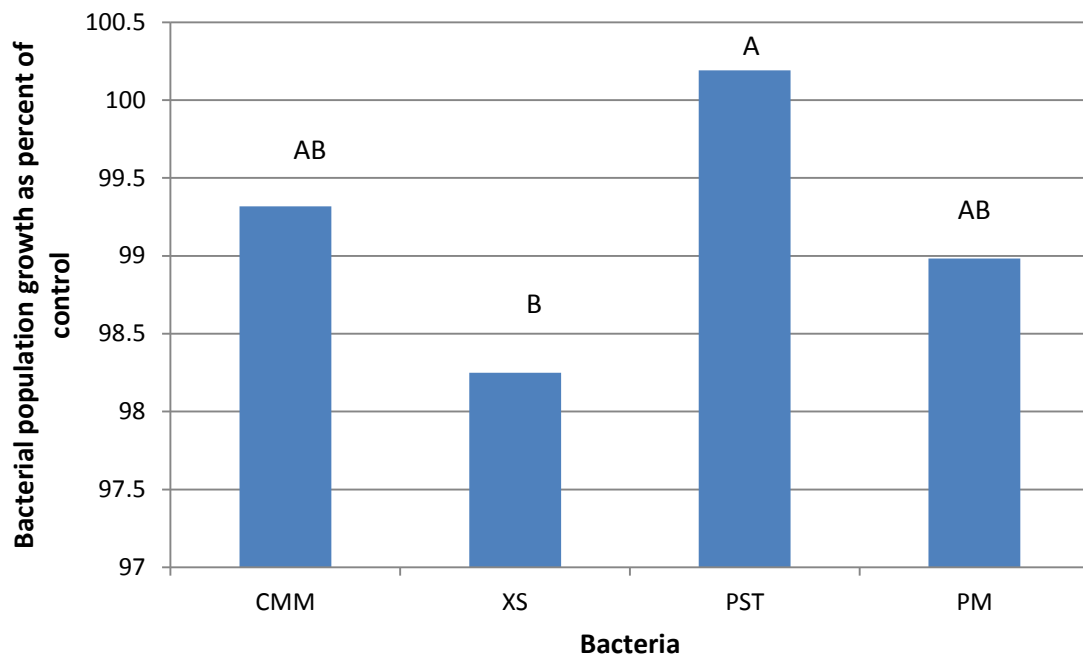


Fig. 1. Effect of bacterial species on percent growth compared to control. CMM = *Clavibacter michiganensis* subsp. *michiganensis*, XS = *Xanthomonas perforans*, PST = *Pseudomonas syringae* pv. *tomato*, PM = *Pseudomonas mediterranea*. Bars with the same letters are not significantly different according to an F-protected LSD at $P = 0.10$.

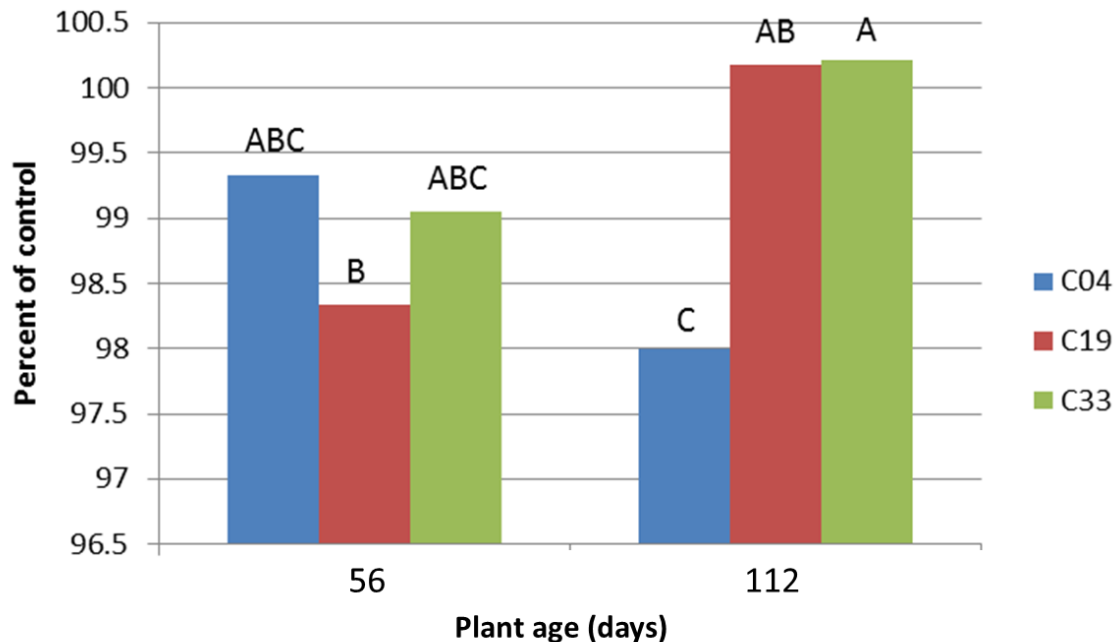


Fig. 2. Effect of the interaction of harvest date $\times$ farm on bacterial population growth as a percent of control. C04, C19, C33 are designations for farms where switchgrass was produced. Bars with the same letters are not significantly different according to an F-protected LSD at $P = 0.05$.

Analysis of the data indicated that significant effects exist. *Xanthomonas* appears to be more susceptible than the other plant pathogens. As the age of plants at harvest increased, the extractives collected from plants on farm C04 had more activity than plants from other locations collected at the same plant age. Since the same cultivar was used, differences in activity of extractives could be due to differences in abiotic and biotic environmental conditions at these locations. Studies of *in vitro* activity of similar switchgrass-derived extractives against food-borne *Salmonella* pathogens showed some activity (Doris D'Souza, personal communication). Due to the environmental stability of the plant

pathogens tested in this study, a longer incubation time may be needed for exposure of the bacteria to the extractives to have a significant effect on bacterial population growth. It is likely that these plant pathogens have evolved to be more tolerant of plant defense compounds than food-borne animal pathogens such as *Salmonella*. Similar compounds have activity against fungal plant pathogens, such as *Fusarium oxysporum*; (Mandal et al. 2009) so extractives may provide a source of antifungal compounds as well as antibacterial compounds. The variations in total phenolics in this study could be further understood with analysis of the specific compounds that make up the total.

References

- Chen, S.F., Mowery, R.A., Scarlata, C.J., and Chambliss, C.K. 2007. Compositional analysis of water-soluble materials in corn stover. J. Agric. Food Chem. 55:5912-5918.
- Chen, S.F., Mowery, R.A., Sevcik, R.S., Scarlata, C.J., and Chambliss, C.K. 2010. Compositional analysis of water-soluble materials in switchgrass. J. Agric. Food Chem. 58:3251-3258.
- Keshwani, D.R., and Cheng, J.J. 2009. Switchgrass for bioethanol and other value-added applications: A review. Bioresource Technol. 100:1515-1523.
- Mandal, S., Mitra, A., and Mallick, N. 2009. Time course study on accumulation of cell wall-bound phenolics and activities of defense enzymes in tomato roots in relation to Fusarium wilt. World J. Microbiol. Biotechnol. 25:795-802.
- Ravindranath, S.V., Uppugundla, N., Lay, J.O., Clausen, E.C., Wilkins, M. Inghraam, R.G., West, C., Wu, Y., and Carrier, D.J. 2009. Policosanol, α -tocopherol, and moisture content as a function of timing of harvest of switchgrass (*Panicum virgatum* L.). J. Agric. Food Chem. 57:3500-3505.
- Tao, L., Aden, A., Elander, R.T., Pallapolu, V.R., Lee, Y.Y., Garlock, R.J., Balan, V., Dale, B.E., Kim, Y.K., Mosier, N.S., Ladish, M.R., Falls, M., Holtzapfel, M.T., Sierra, R., Shi, J., Ebrik, M.A., Redmond, T., Yang, B., Wyman, C.E., Hames, B., Thomas, S. and Waner, R.E. 2011. Process and technoeconomic analysis of leading pretreatment technologies for

- lignocellulosic ethanol production using switchgrass. *Bioresource Technol.*
doi: 10.1016/j.biortech.2011.07.051.
- Thammasouk, K., Tandjo, D., and Penner, M.H. 1997. Influence of extractives on
the analysis of herbaceous biomass. *J. Agric. Food Chem.* 45:437-443.
- Uppugundla, N., Engelberth, A., Ravindranath, S.V., Clausen, E.C., Lay, J.O.,
Gidden, J., and Carrier, D.J. 2009. Switchgrass water extracts: Extraction,
separation and biological activity of rutin and quercitrin. *J. Agric. Food
Chem.* 57:7763-7770.
- Waterhouse, A.L. 2002. Determination of total phenolics. *Current Protocols in
Food Analytical Chemistry* 1.1.1-1.1.8.
- Yan, J., Hu, Z., Pu, Y., Brummer, E.C., and Ragauskas, A.J. 2010. Chemical
compositions of four switchgrass populations. *Biomass and Bioenergy.*
34:48-53.

CHAPTER 5

CONCLUSION

The results of this project provide information that will be integral for the development of disease management practices and seed certification programs for switchgrass in Tennessee. Nine fungal pathogens were identified from mature switchgrass grown in Tennessee, and at least five fungal pathogens were isolated directly from switchgrass seed produced in the United States and shipped to Tennessee. As the biofuels initiative at the University of Tennessee moves forward, disease management practices and seed certification programs could significantly increase both the commercial and overall success of the switchgrass-derived cellulosic ethanol program. Further studies into the effect of fungal infection on extractives composition, fermentable sugar yield per gram biomass, and total cellulosic ethanol yield per gram biomass will need to be conducted to determine the overall effect of fungal infection on the cellulosic ethanol system of switchgrass. The ethanol-soluble extractives of switchgrass show some antibacterial activity, and further studies will be needed to determine optimal extractives concentration and treatment conditions, as well tests for antifungal activity and economic and energy-expenditure studies for the feasibility of removing ethanol-soluble extractives prior to the pretreatment stage of the biomass conversion process of switchgrass.

VITA

Andrea Vu grew up in Pembroke, Massachusetts. She received a Bachelor of Science degree in Plant and Soil Science from the University of Massachusetts at Amherst in May 2008. After taking a small amount of time off from academics, Andrea moved to Knoxville, TN to pursue further education in Plant Pathology. In December 2011, she earned a Master of Science degree in Entomology and Plant Pathology under the direction of Dr. Bonnie Ownley.